

PIPSA

Comparison of Protein Interaction Properties

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<http://www.eml-research.de/mcm>

SYCAMORE: Systems biology's Computational Analysis and Modeling Research Environment

<http://sycamore.eml.org>

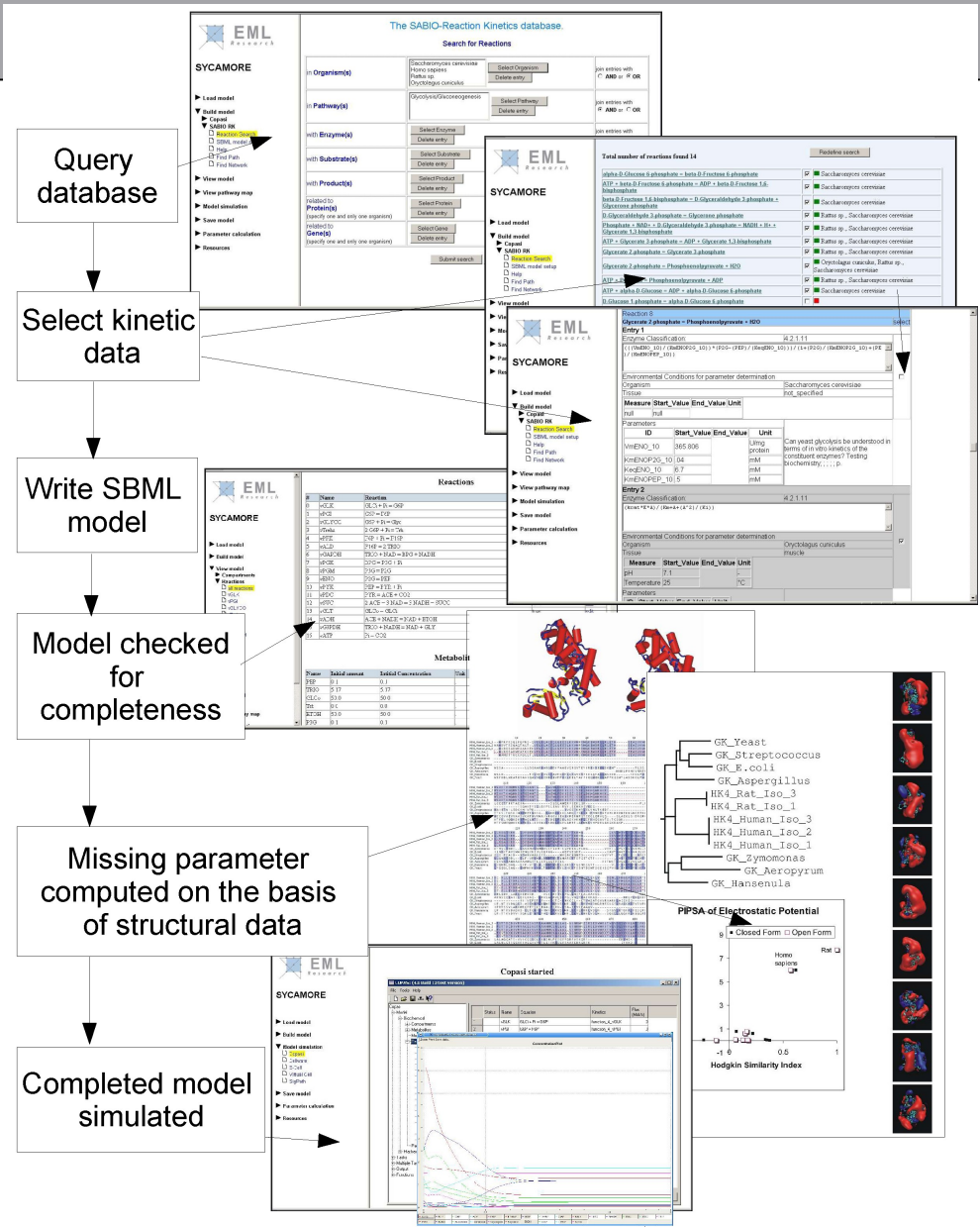
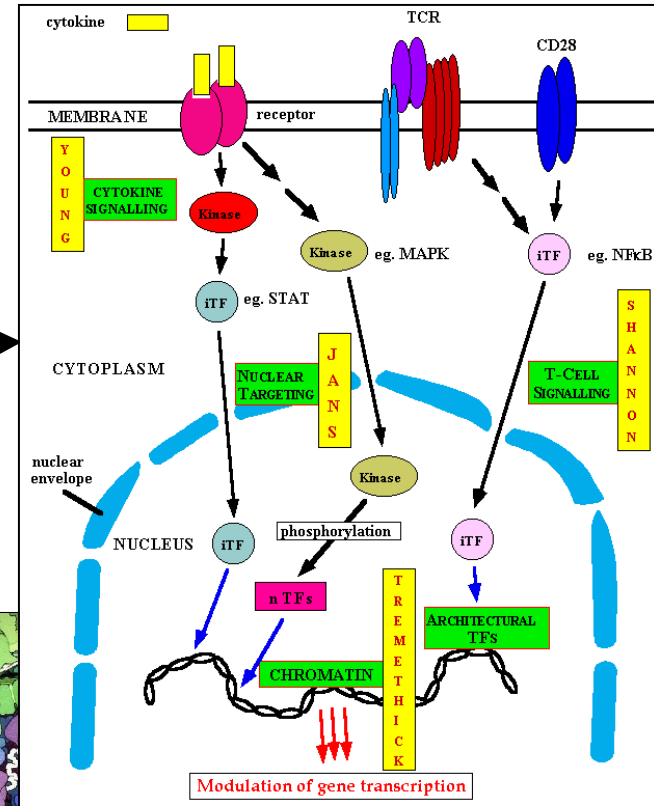
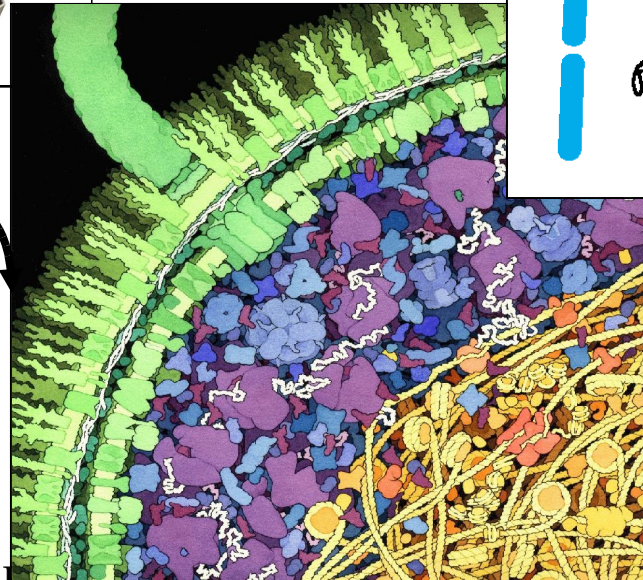
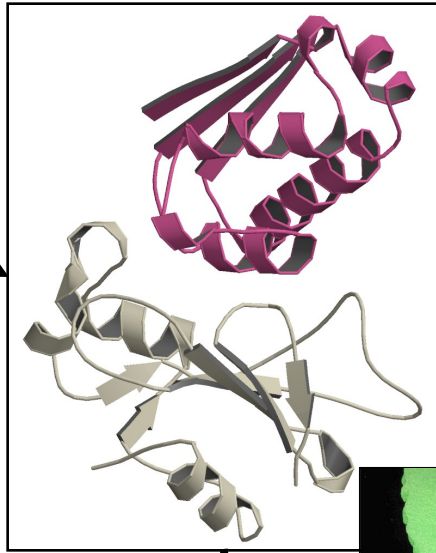
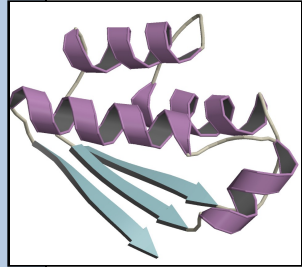


Fig 1: Example of using the available features in SYCAMORE. The case study is modeling and simulating glycolysis in hepatocytes. First, the database is queried and the relevant kinetic data selected. Then the SBML model file is created. This is checked for completeness (this is not implemented for automatic use yet). A missing parameter (here we assume Km for glucokinase to be missing) is then computed using structural data. Finally, the completed model is simulated.

- introduction
- protein structures
- molecular interaction fields
- electrostatic potentials
- PIPSA
- webPIPSA
- visualisation and analysis
- qPIPSA
- tutorial with examples

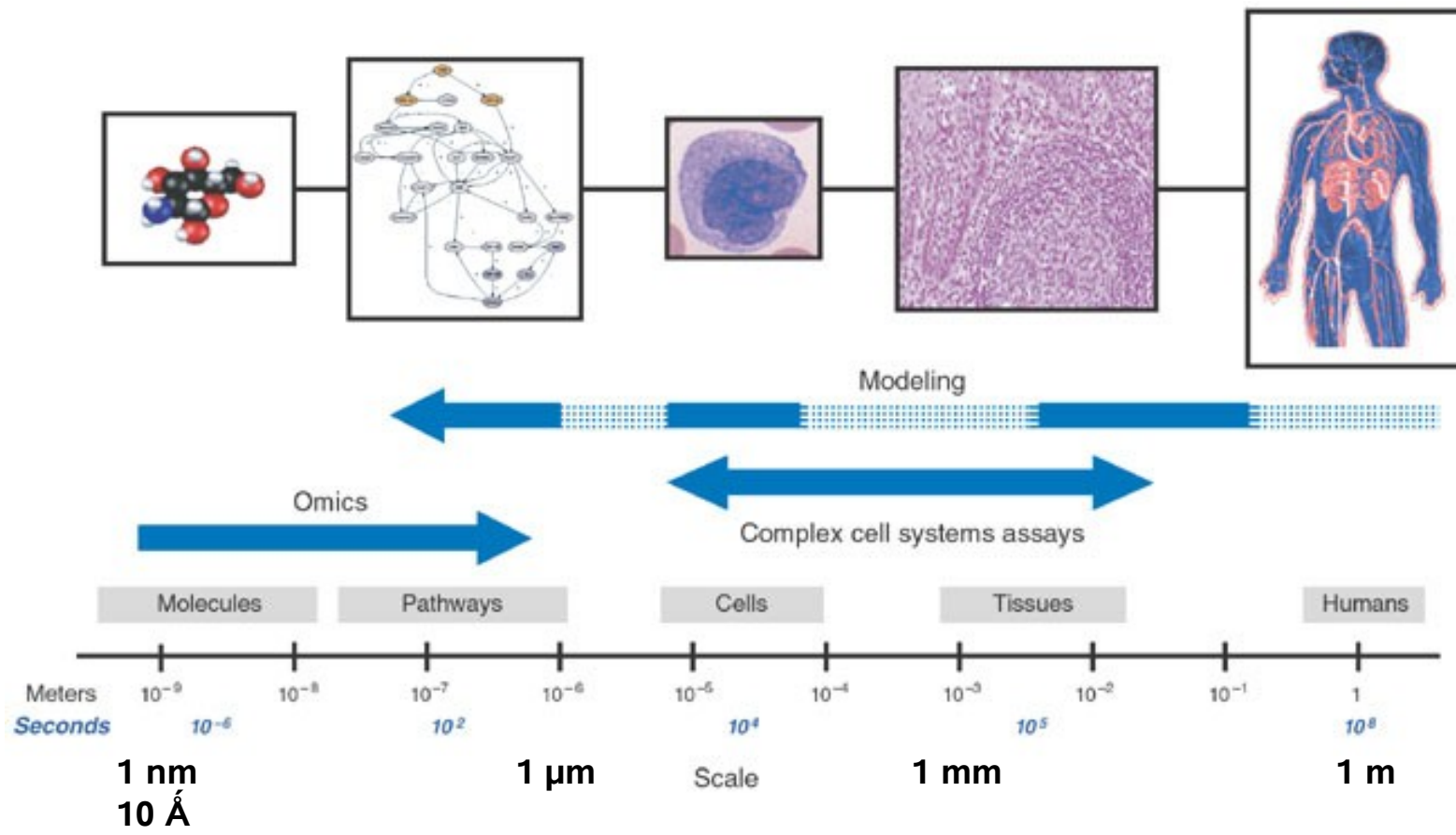
Introduction

From Individual Molecules to Cellular Systems



David Goodsell
in Voet,
Biochemie, 1998

dimensions



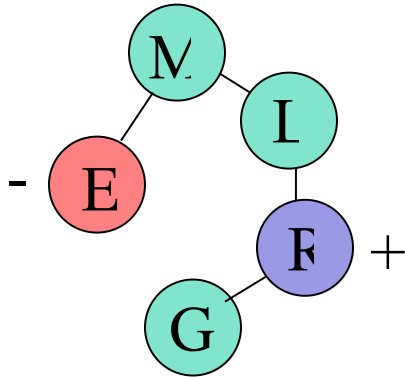
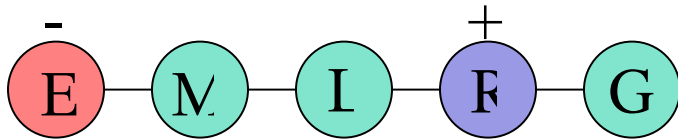
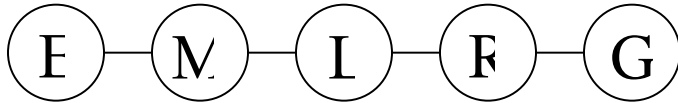
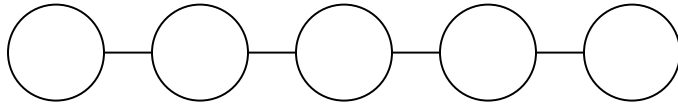
20
natural
amino
acids

Sidechain polarity:

- Non-polar
- Polar
- negatively charged
- positively charged

Alanine	Ala	A	(Alanine)
Cysteine*	Cys	C	(Cysteine)
Aspartic Acid	Asp	D	(aciD)
Glutamic Acid	Glu	E	(E comes after D)
Phenylalanine	Phe	F	(Ph=F)
Glycine**	Gly	G	(Glycine)
Histidine***	His	H	(Histidine)
Isoleucine	Ile	I	(Isoleucine)
Lysine	Lys	K	(L follows K)
Leucine	Leu	L	(Leucine)
Methionine	Met	M	(Methionine)
Asparagine	Asn	N	(AsparagiNe)
Proline	Pro	P	(Proline)
Glutamine	Gln	Q	(Qlutamine)
Arginine	Arg	R	(aRginine)
Serine	Ser	S	(Serine)
Threonine	Thr	T	(Threonine)
Valine	Val	V	(Valine)
Tryptophan	Trp	W	(Double ring - W)
Tyrosine	Tyr	Y	(tYrosine)

protein folding



20
natural
amino
acids

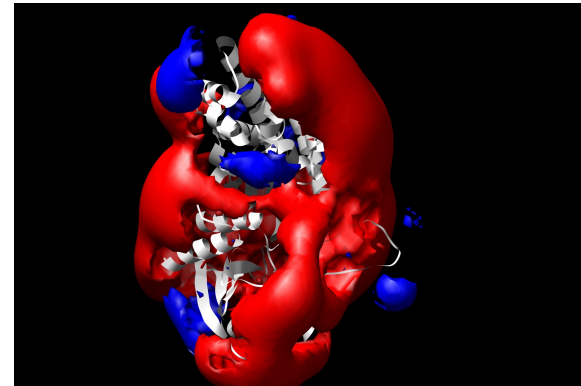
E = Glutamic Acid

M = Methionine

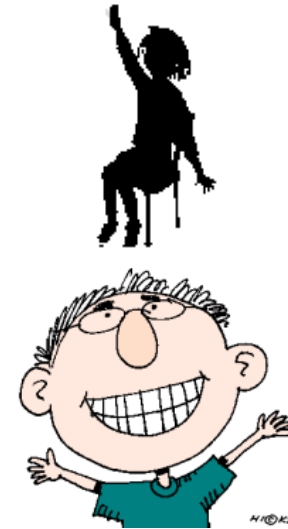
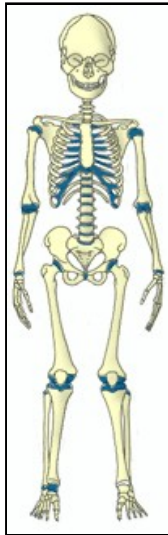
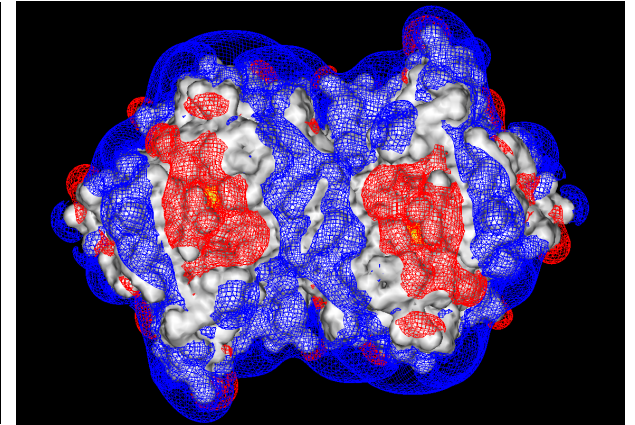
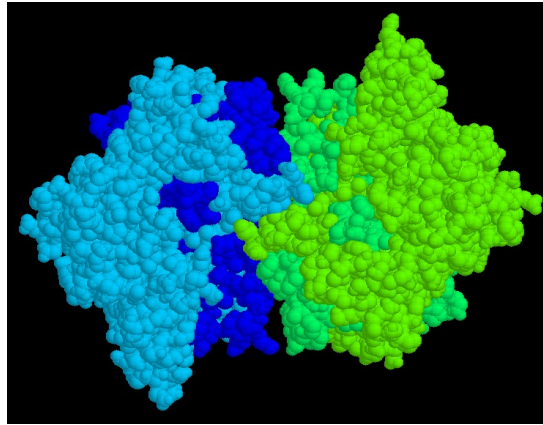
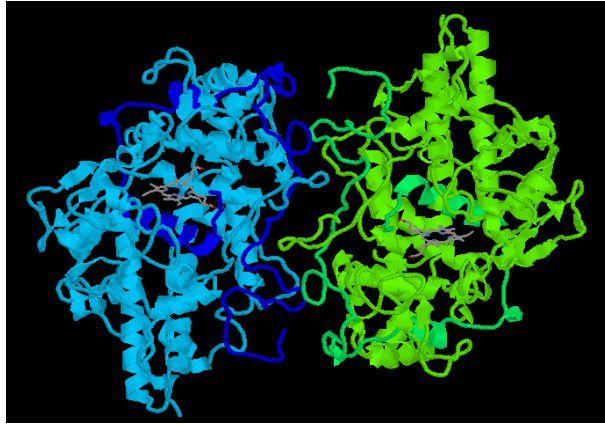
L = Lysine

R = Arginine

G = Glycine

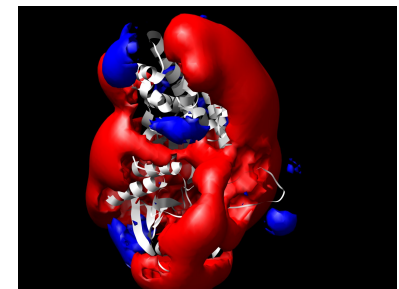
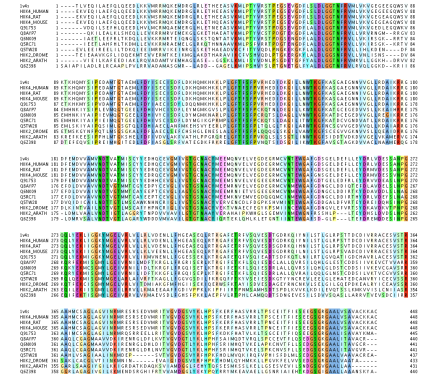


protein flexibility and biomolecular recognition



levels of protein comparison

- properties
 - mass, isoelectric point, charge, fingerprints,...
- amino acid sequence
 - bioinformatics (sequence alignment, prediction of motifs)
- protein structures
 - X-ray, NMR, EM
- protein structure/function relationship
 - QSAR
 - molecular interaction fields (MIFs)



Exploring Protein Interactions via the Computer

- Small molecules
- Proteins
- Nucleic acids
-

Protein Interactions

- Stability
- Folding
- Catalysis
- Electron transfer
-

- Physics-based
- Bio/Chemo-Informatics

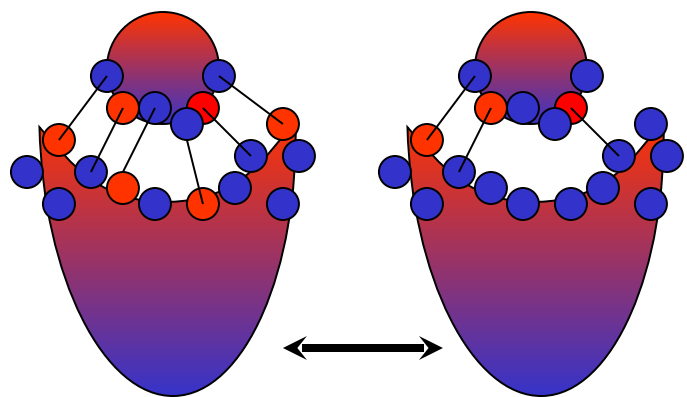
Methods

Applications

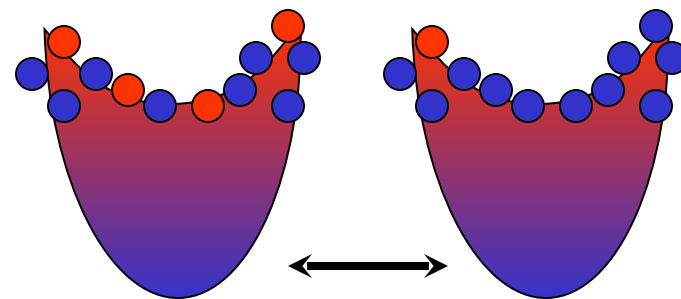
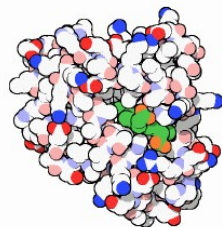
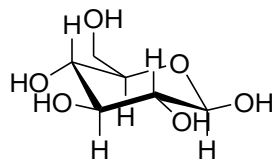
- Individual proteins – in-depth
- Proteins “in context” – biochemical pathways
- Comparison across large protein families

Molecular Interaction Fields

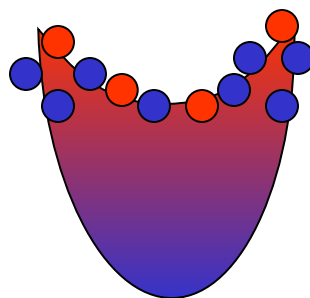
Comparing Molecular Interaction Fields



comparing interactions

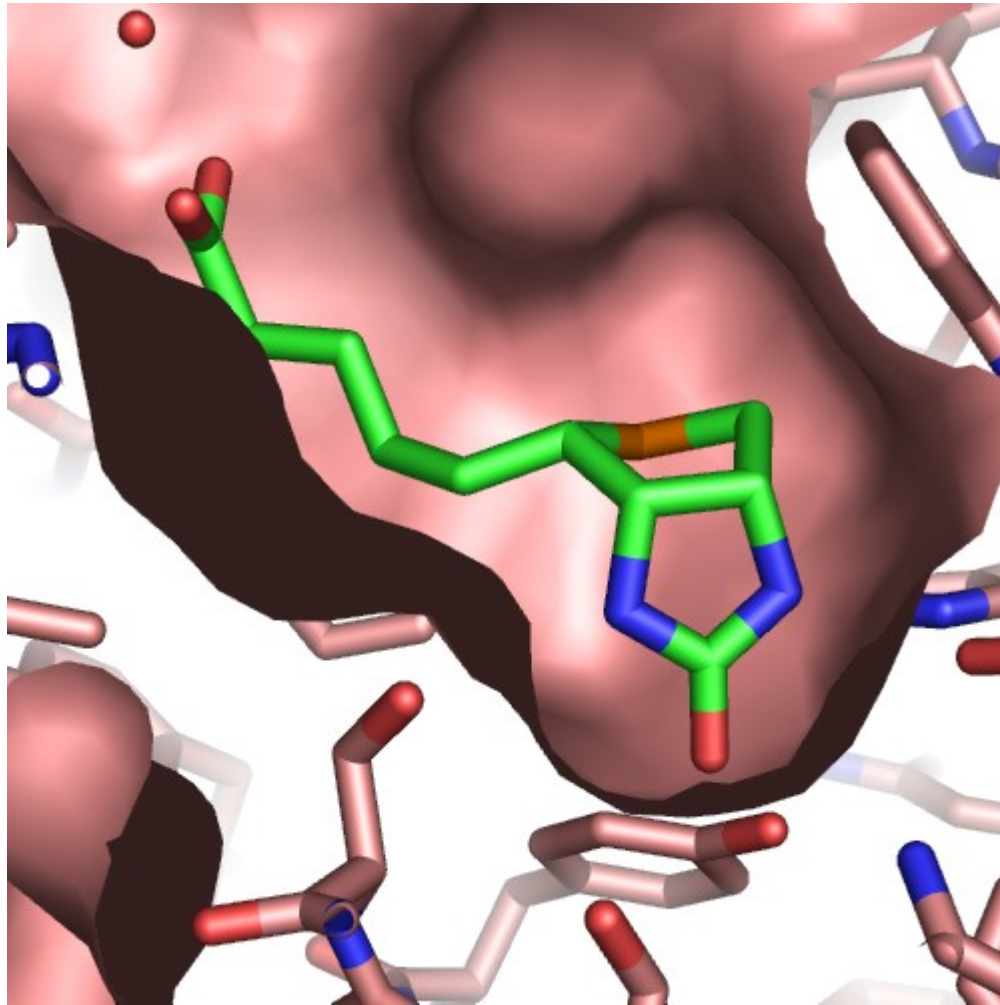


comparing interaction fields

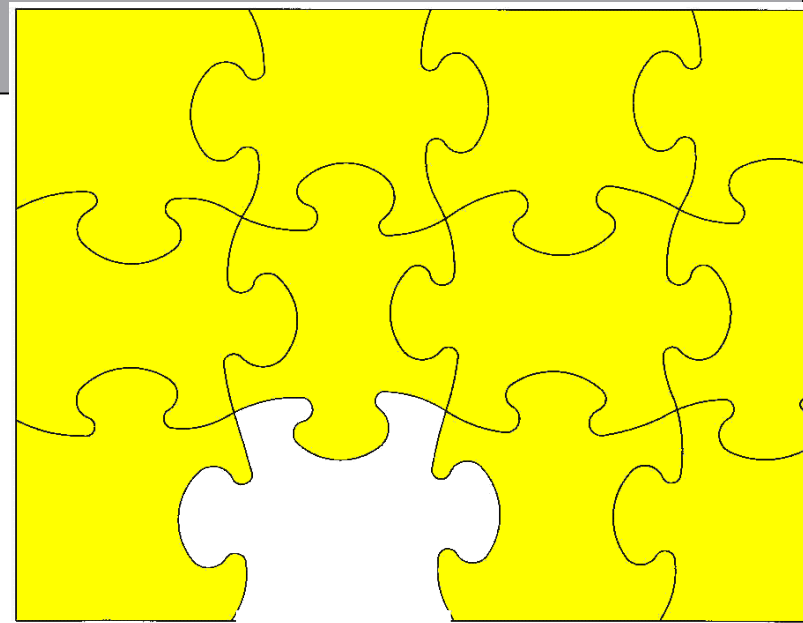


protein-ligand
protein-protein

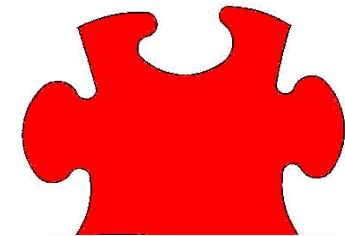
- Charge distribution
- Potential
- Shape
- ...



Streptavidin complex with biotin (1stp)



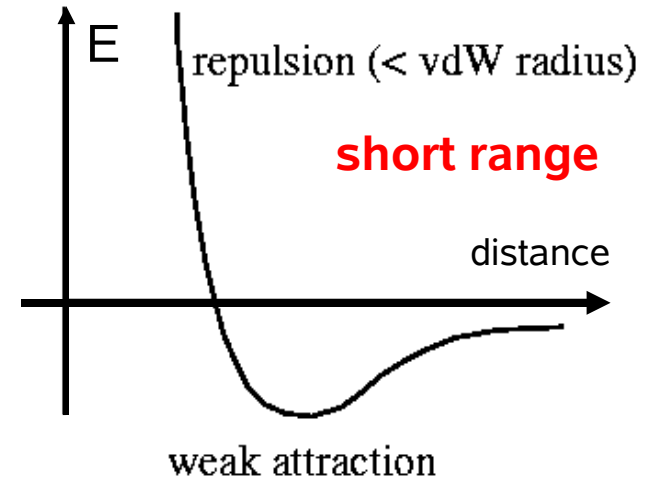
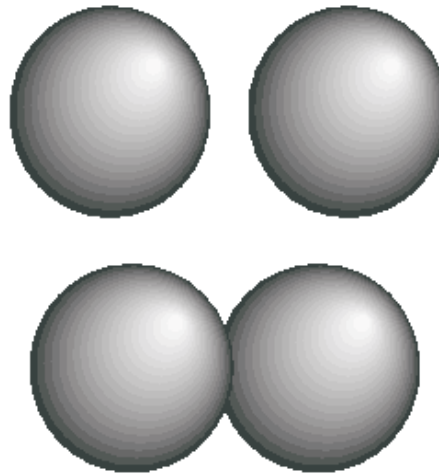
**shape
complementarity**



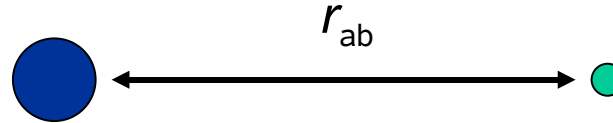
non-bonded interactions

Lennard-Jones (van der Waals)

$$E_{LJ} = \frac{A}{r_{ab}^{12}} - \frac{B}{r_{ab}^6}$$

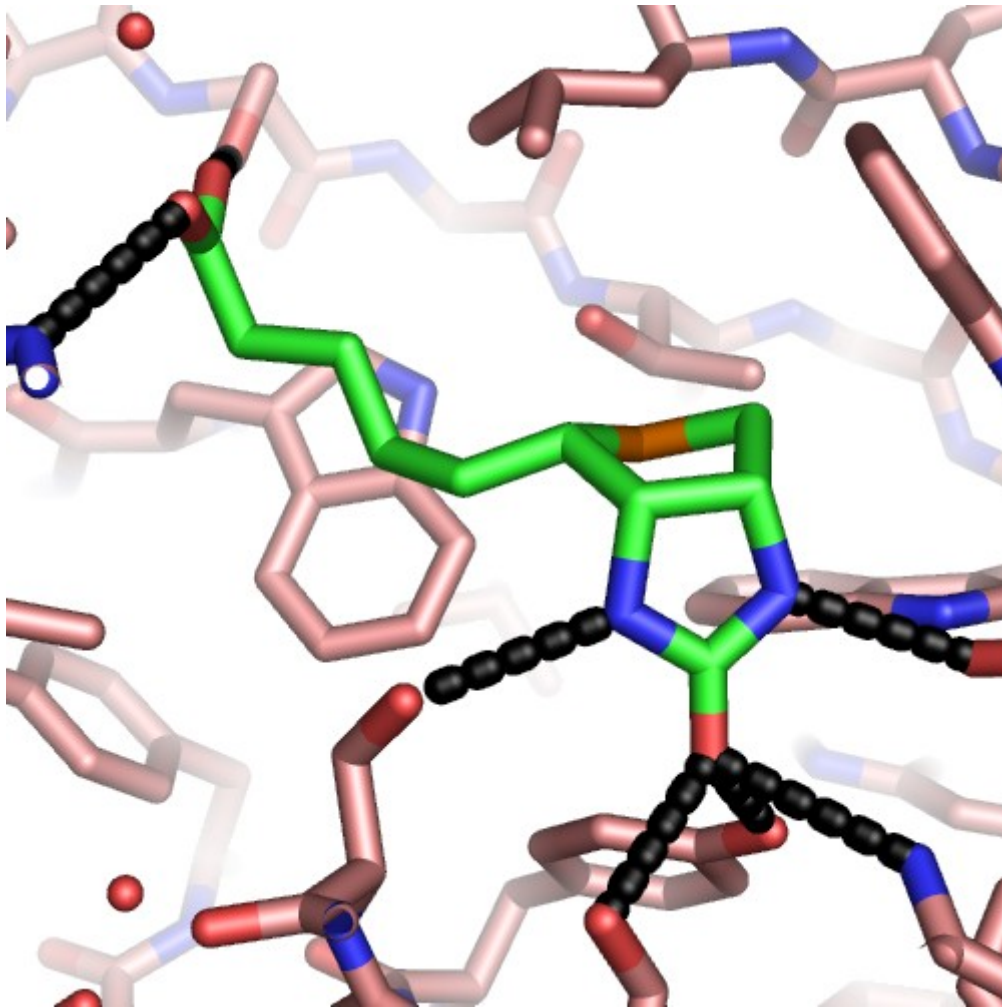


$A = 0.5 C (R_a + R_b)^6$
C: Slater-Kirkwood formula
 R_a and R_b : van der Waals radii



- van der Waals excluded volume repulsion
- London dispersive induced-dipole induced-dipole attraction
- **short-range** (8-10 Å cutoff)

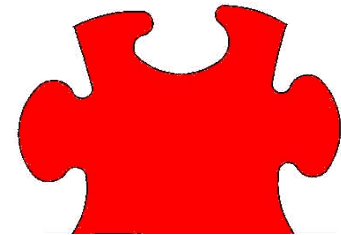
receptor-ligand binding



Streptavidin complex with biotin (1stp)



**electrostatic
complementarity**



Coulomb's Law

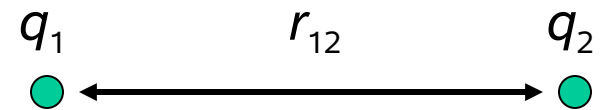
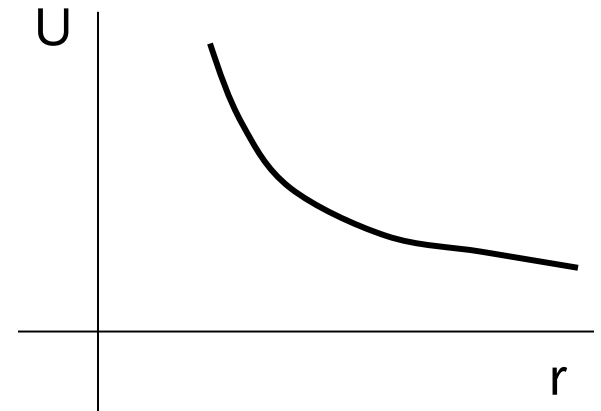
- interaction energy of two point-charges *in vacuo*
- solve Poisson equation
- in SI units:

$$U = \frac{q_1 q_2}{4\pi\epsilon_0 r_{12}}$$

ϵ_0 = permittivity of free space
($\epsilon_0 = 8.85 \cdot 10^{-12} \text{ F m}^{-1}$)

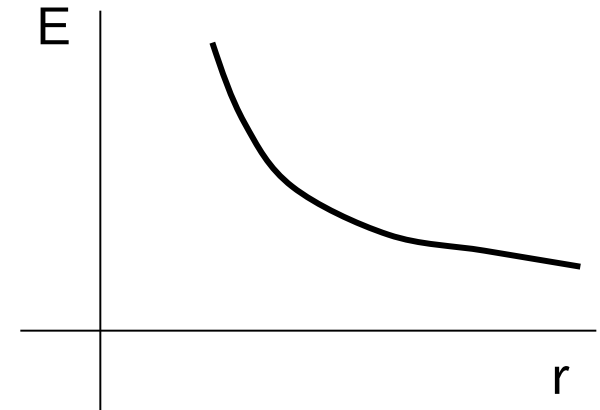
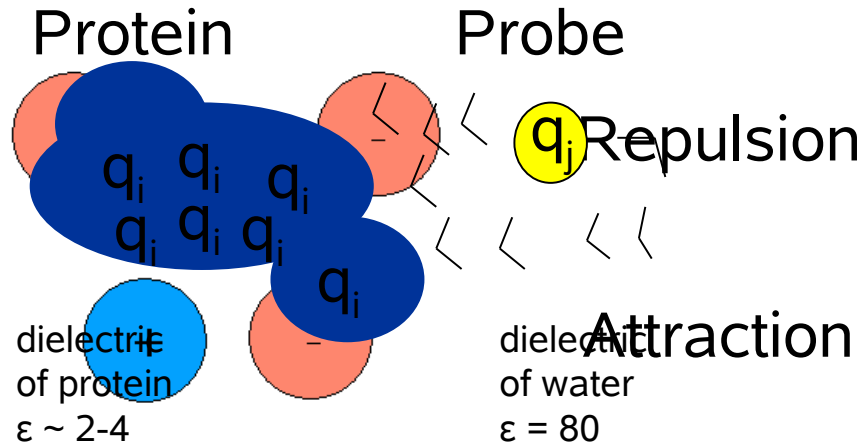
- in “Biomolecular” units:

$$U = \frac{332q_1q_2}{r_{12}}$$



Energy, U: kcal/mol
Charge, q: electron charges
Distance, r: Ångstroms

Non-bonded interactions: Electrostatic



$$E_{ele} = \frac{q_i q_j}{\epsilon \pi \epsilon_0 \epsilon_r r_{ij}}$$

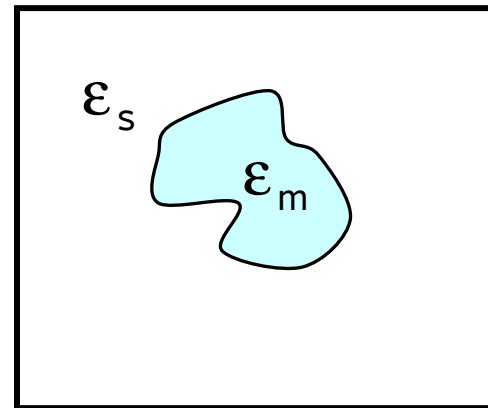


- Coulombic forces
- Each atom has partial atomic charge q_1, q_2
- Compared to vacuum, the uniform medium weakens (damps/screens/shields) electrostatic interactions between charges
- ϵ_0 : permittivity of free space,
 ϵ_r : relative dielectric constant of the surrounding medium
- **Long range** ($\sim 1/r$) (No cutoff)

- Electronic
 - Electrons and nuclei of an atom are shifted in opposite directions by the external field
 - Linearly proportional to applied field
 - $\epsilon_r \sim 1.5-2.5$
- Orientational
 - Molecules that have a permanent dipole moment, e.g. water ($\mu_D = 1.9 \text{ D}$), align to oppose the external field
 - Average dipole moment rises linearly at weak fields, saturates at high fields
 - ϵ_r up to ca. 80 (water)

Continuum electrostatics for molecules

- dielectric constant is a macroscopic property
- nevertheless, it can be applied microscopically
 - Molecule
 - Part of molecule
- for small molecules, e.g. ethanol, polarizability is primarily electronic
 - Molecule of $\epsilon_M \sim 1.5-2.5$
surrounded by vacuum of $\epsilon_s \sim 1$ or water of $\epsilon_s \sim 80$ etc
- for macromolecules, orientational polarizability is also important
 - $\epsilon_M \sim 2-80$, dependent on
 - location
 - theoretical model
- non-uniform dielectric
 - Dielectric boundaries



Effective dielectric constant

- non-uniform dielectric leads to distortion of field due to a point charge
- due to total dielectric environment
- effects such as electrostatic focusing

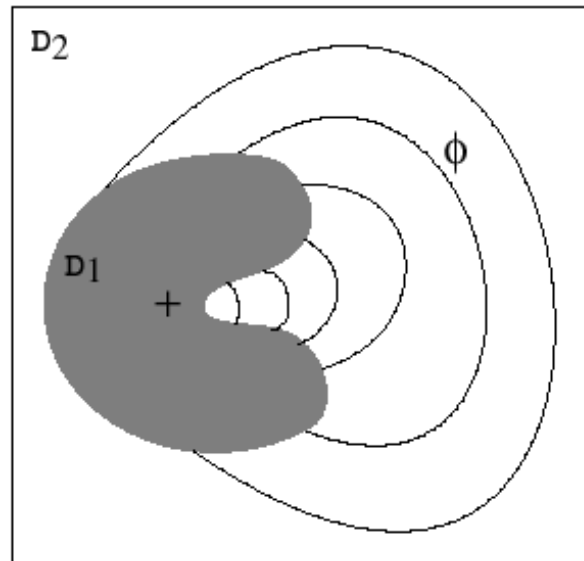


Figure: M. K. Gilson

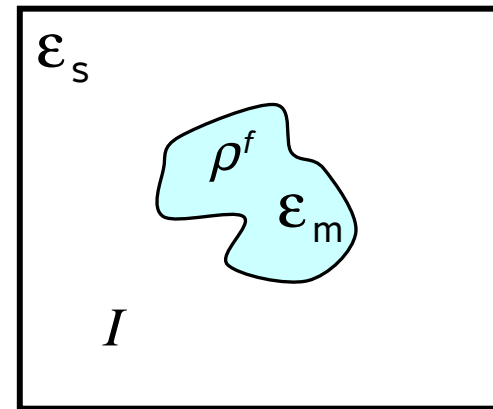
- Linearized Poisson-Boltzmann equation

$$-\varepsilon_0 \nabla \cdot [\varepsilon_r(\mathbf{r}) \nabla \phi(\mathbf{r})] = \rho^f(r) - \varepsilon_0 \varepsilon_r(r) \kappa^2(r) \phi(r)$$

$$\kappa^2(r) = \frac{\beta}{\varepsilon_0 \varepsilon_r} \sum_1^N c_{i,bulk} q_i^2 = \frac{2e^2 N_A}{\varepsilon_0 \varepsilon_r kT} I$$

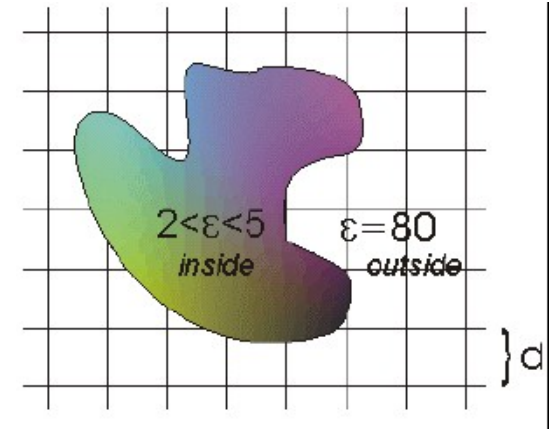
- Finite-difference
- Numerical solution

$\phi(\mathbf{r})$ =Elec. Potential
 $\rho(\mathbf{r})$ =Charge density



Solving the Poisson-Boltzmann equation

- Analytic solutions
 - Only for simple geometries: spheres, planar boundaries
- Numerical solutions
 - Convert continuous partial differential equation into discretized problem with systems of linear equations that can be solved by matrix methods



Finite difference method

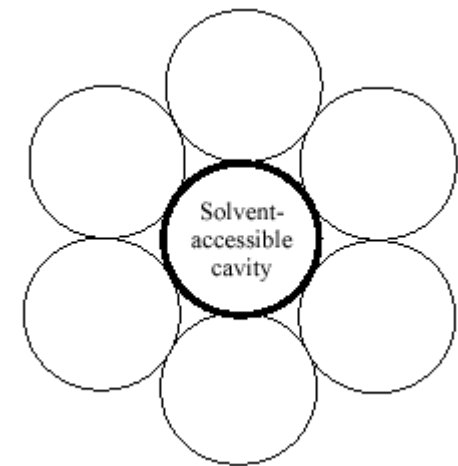
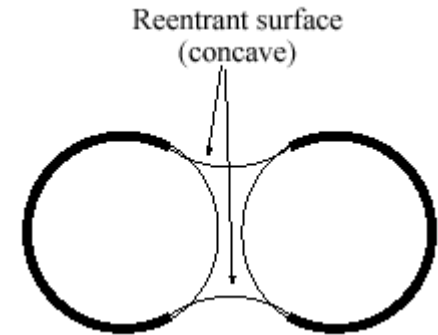
- Discretize space into a cubic lattice, solve iteratively to convergence for each grid point by finite differences

Solving the finite difference linearized Poisson-Boltzmann equation: Practical aspects

- Need to assign:
- Atomic radii and charges
 - From “standard” force field e.g. CHARMM, AMBER
 - Set specifically for continuum electrostatics, parameterized to reproduce small molecule solvation energies
- Grid size and spacing
 - Large enough for boundary potentials to be sufficiently accurate
 - Focusing using nested grids is possible
 - Spacing of 0.5-1 Å for viewing potentials or Brownian dynamics forces
 - Spacing of 0.2-0.3 Å for energies, pK_a s

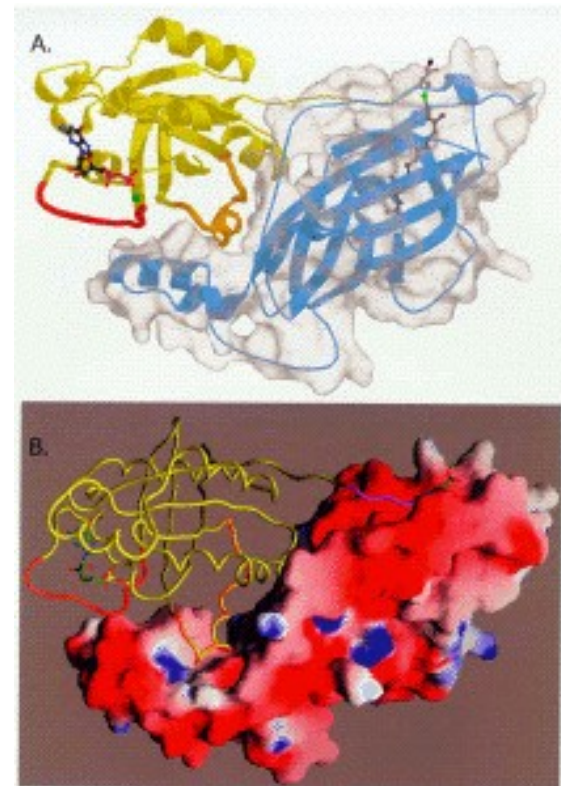
Solving the finite difference linearized Poisson-Boltzmann equation: Practical aspects

- Need to assign:
- Dielectric boundaries
 - Van der Waals surface
 - Molecular surface
 - Smooth dielectric constant over points adjacent to the boundary

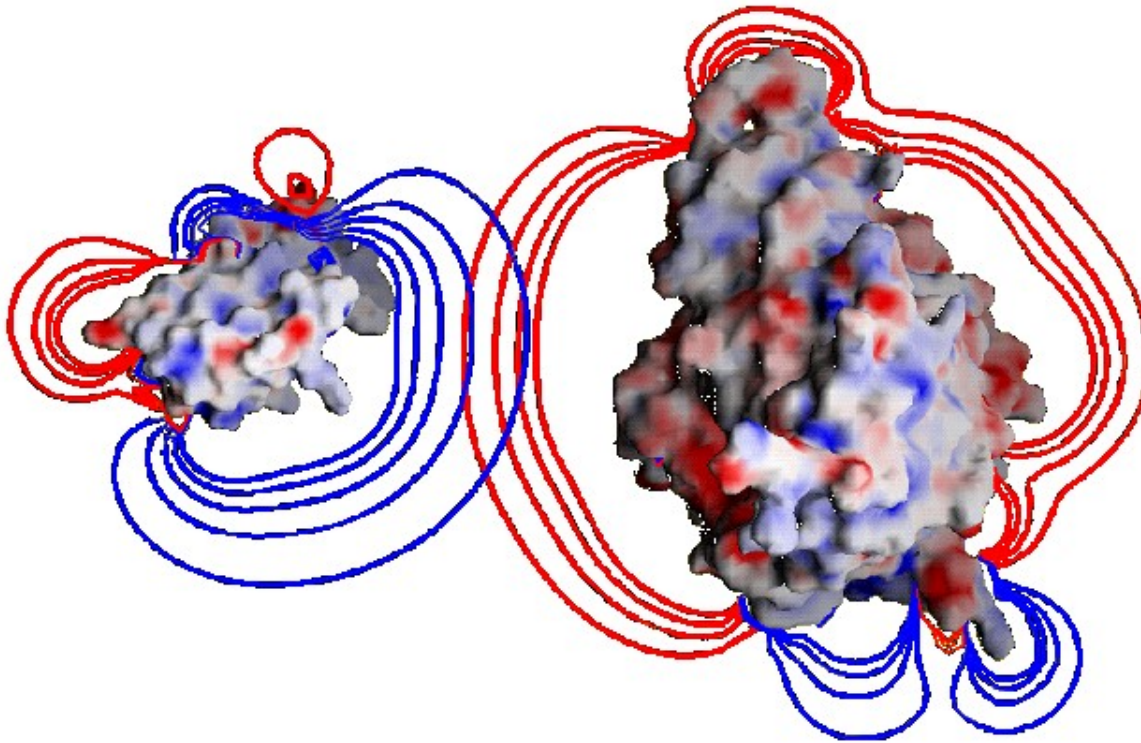


Figures: M. K. Gilson

- Electrostatic potentials
 - Binding complementarity
 - Molecular multipoles
 - Visualization
 - Protein Interaction Property Similarity Analysis (PIPSA)

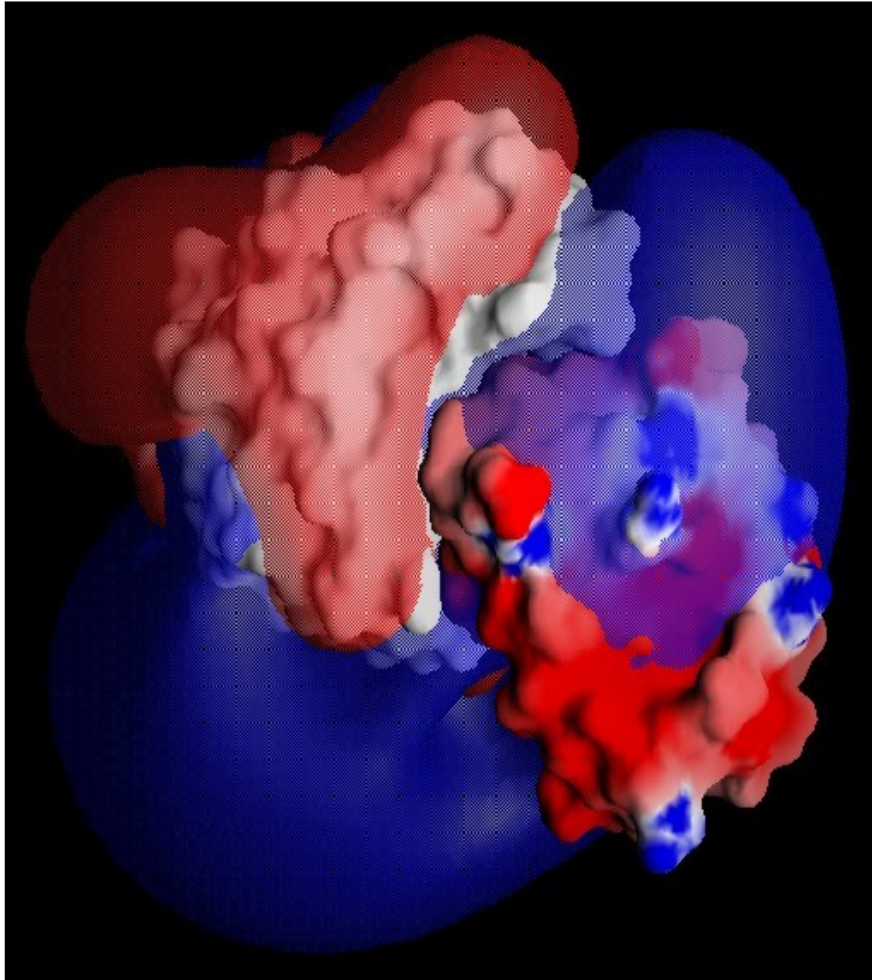


Fasciculin-Acetylcholinesterase: Electrostatic Complementarity

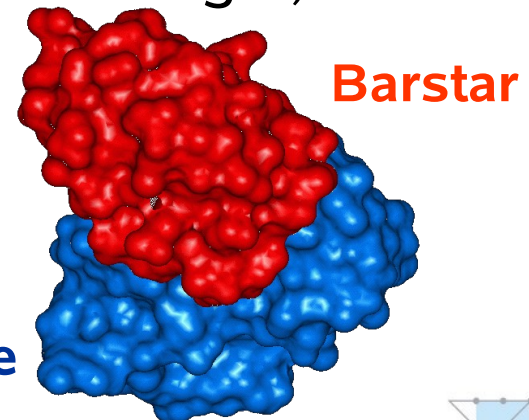


- AChE: $-6e$
 - neurotransmitter hydrolysis
 - cholinergic synapses
- Fasciculin: $+4e$
 - neurotoxin inhibitor
- $K_d \sim 10^{-13} \text{ M}$
- $K_{on} \sim 10^9 \text{ M}^{-1}\text{s}^{-1}$
(zero ionic strength)

Barnase-Barstar: Electrostatic Complementarity

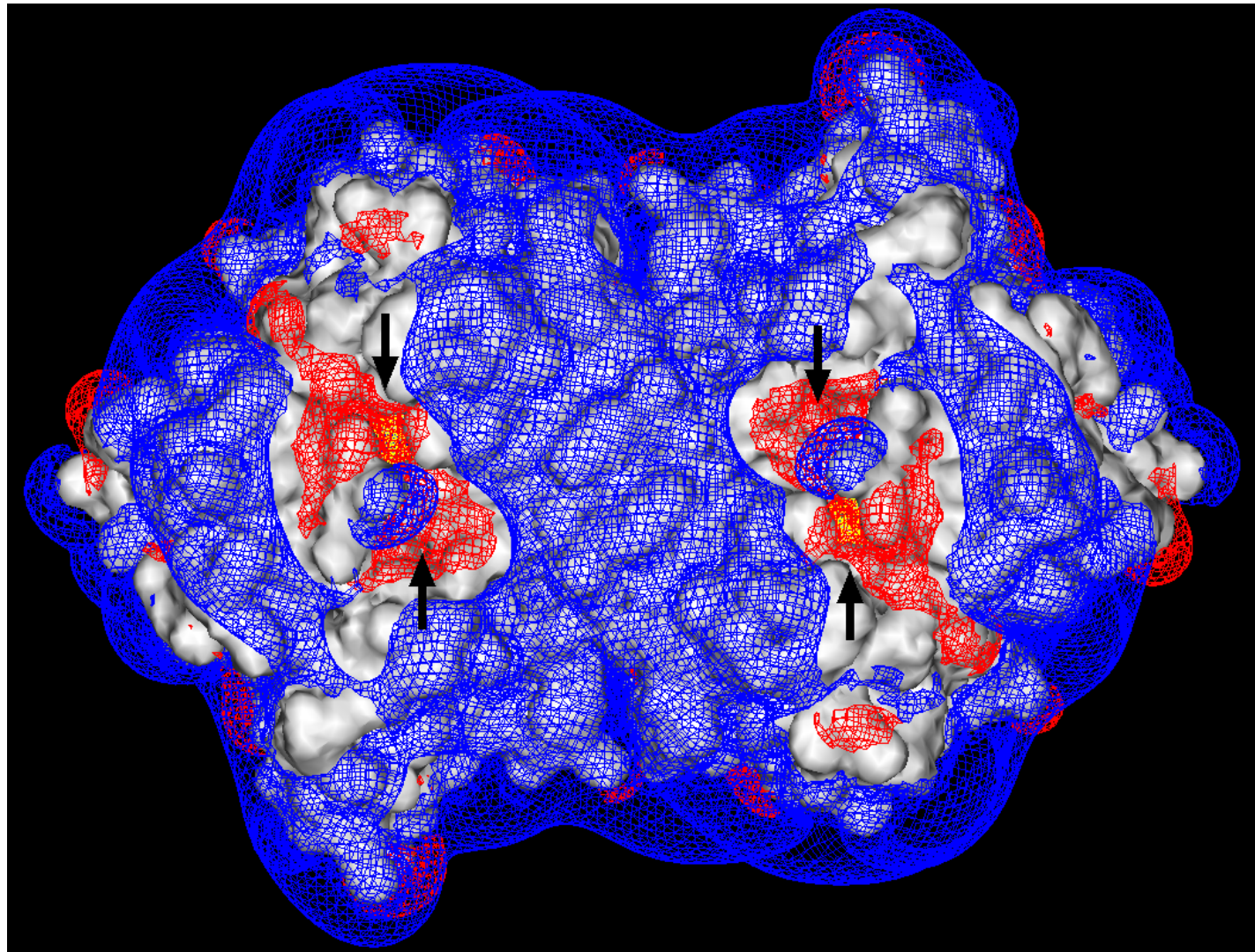


- Barnase : +2e
 - extracellular ribonuclease
 - *Bacillus amyloliquefaciens*
- Barstar : – 6e
 - intracellular inhibitor
- $K_d \sim 10^{-14} \text{ M}$
- $K_{on} \sim 10^{10} \text{ M}^{-1}\text{s}^{-1}$ (zero ionic strength)



Barnase

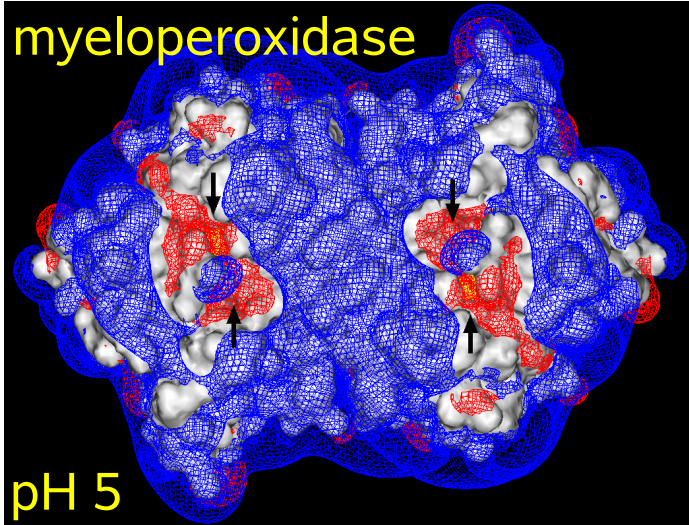
Myeloperoxidase: Active site access



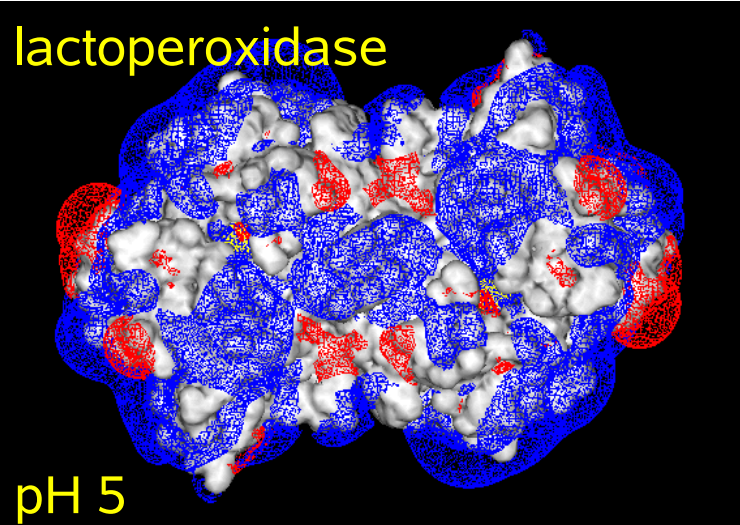
Gabdoulline, Kummer, Olsen, Wade, Biophys. J (2003) 85, 1421-1428

Electrostatics of 3 Peroxidases

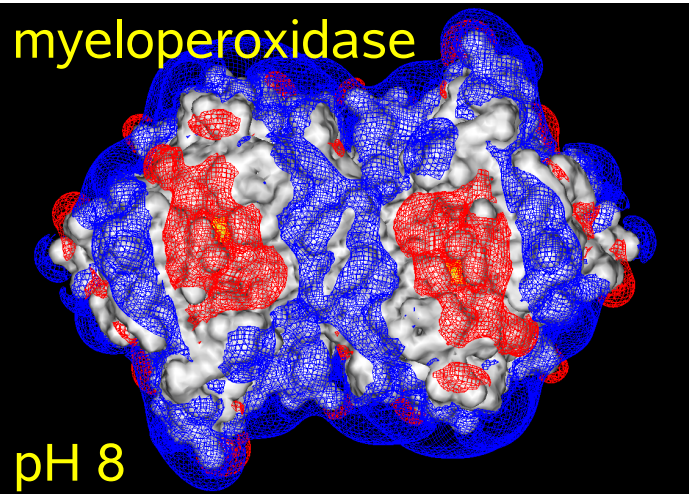
myeloperoxidase



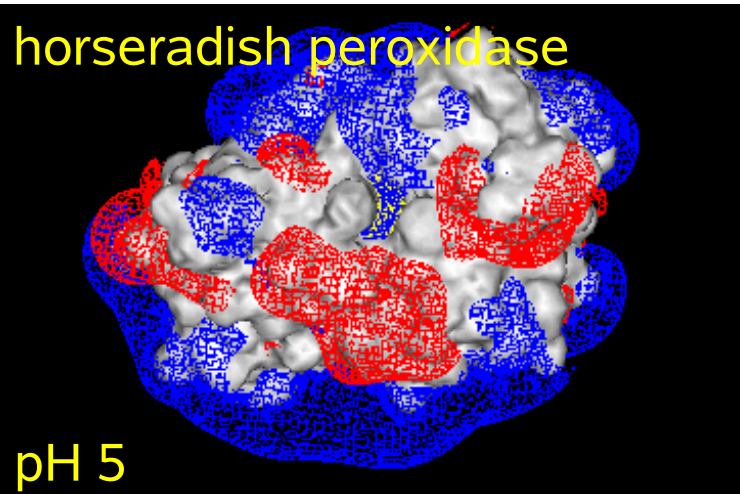
lactoperoxidase



myeloperoxidase



horseradish peroxidase



University of Houston Brownian Dynamics (UHBD)

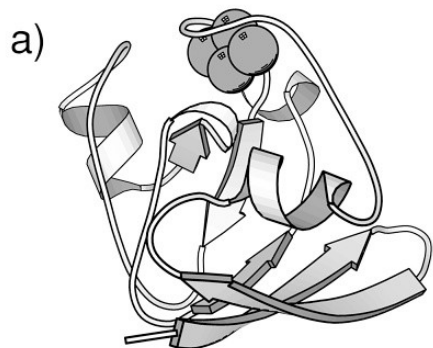
- solves the linearized and non-linearized Poisson-Boltzmann equation

<http://adrik.bchs.uh.edu/uabd.html>

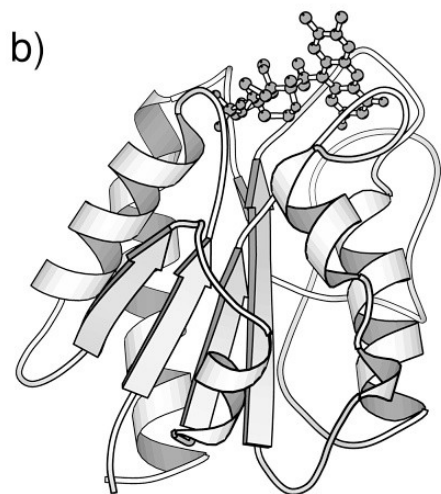
Adaptive Poisson-Boltzmann Solver (APBS)

<http://apbs.sourceforge.net>

Structural Alignment based on Electrostatic Potential

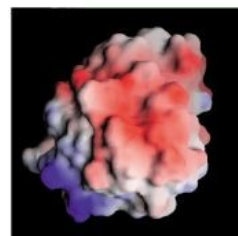


G. M. Ullmann,
M. Hauswald,
A. Jensen,
E. W. Knapp
Proteins 2000

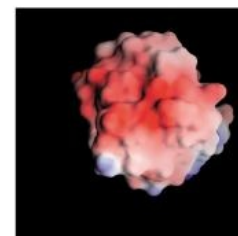


Structures of ferredoxin and flavodoxin.
Although both proteins differ in structure and size, they perform the same physiological function.

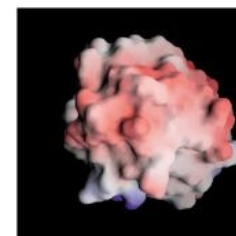
Flavodoxin
(Alignment 1)



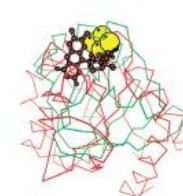
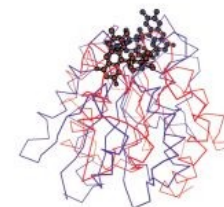
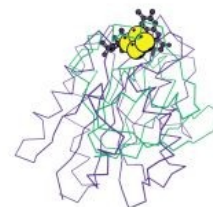
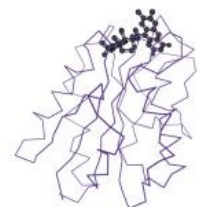
Ferredoxin



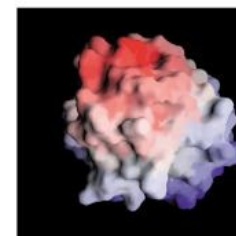
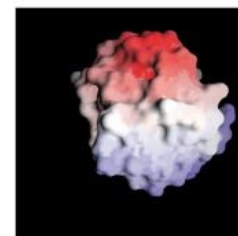
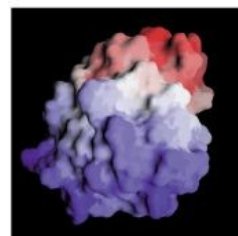
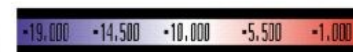
Flavodoxin
(Alignment 2)



Electrostatic Potential



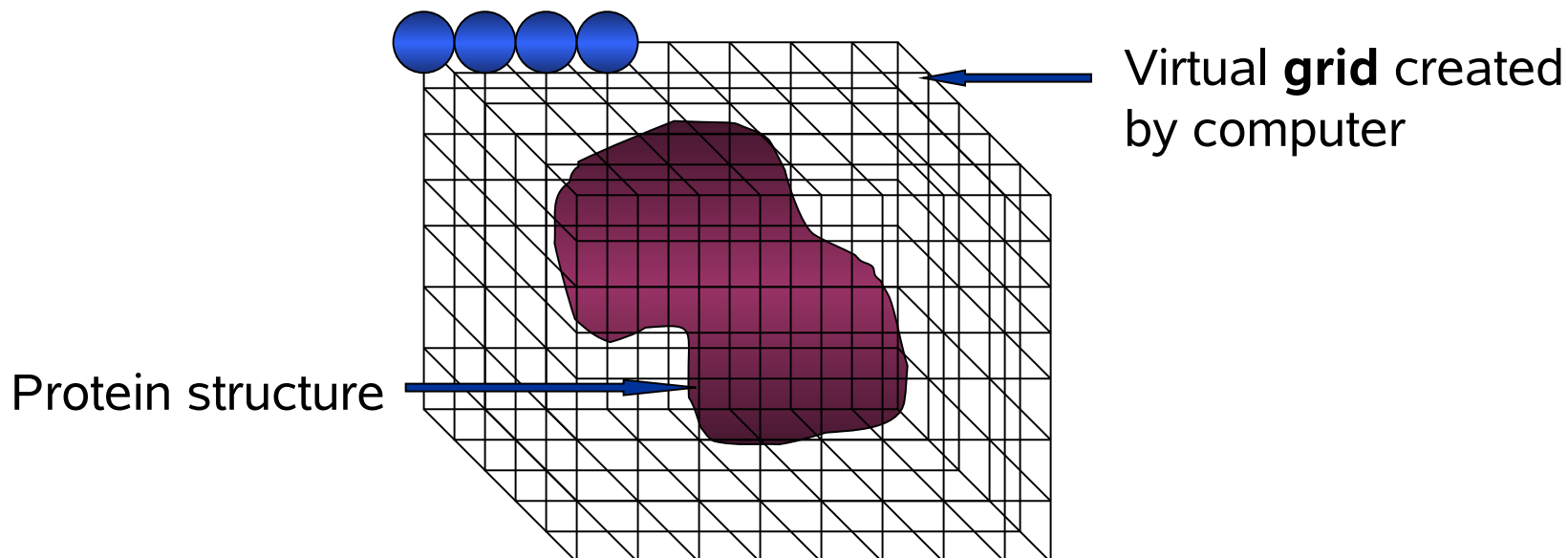
Relative Electronic Coupling



GRID

other examples for
molecular interaction fields (MIFs)

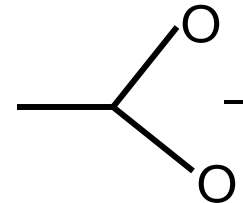
- Detect energetically favorable binding sites on molecules of known structure



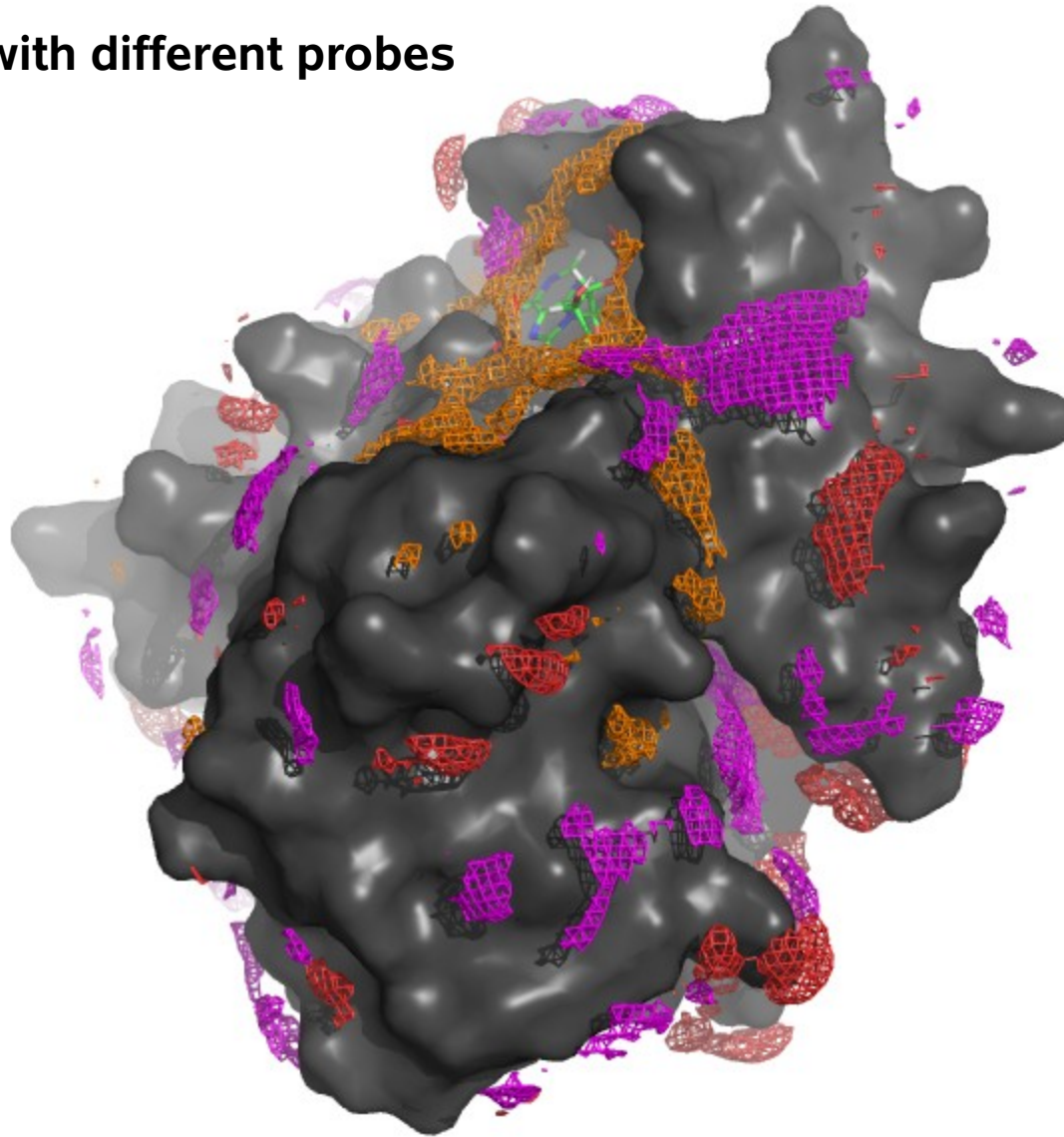
$$\Delta E = \sum_i E_{LJ} + \sum_i E_{EL} + \sum_i E_{HB} + S$$

Goodford, PJ *J. Med. Chem.* (1985) 28, 849-857.

- Most are appropriately parameterized single-point spheres
 - E.g. carbonyl oxygen, hydroxyl, amino, water, dry, sulfate
- Some are multiatom
 - E.g. carboxylate, amide
- Water Probe
 - Hydrophilic protein surface
- Dry Probe
 - Hydrophobic protein surface

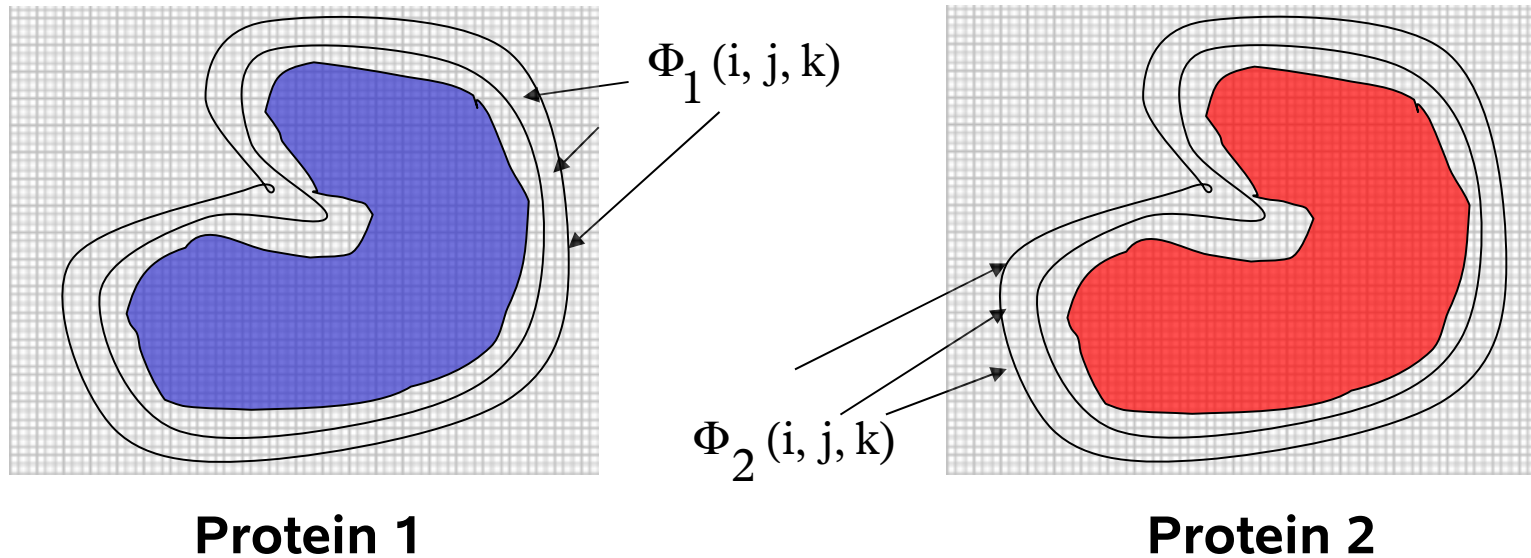


using Grid with different probes



PIPSA

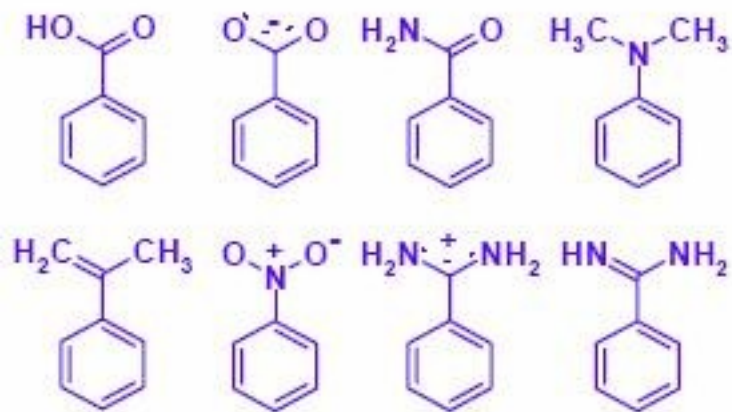
- Classification of the proteins in a protein family by interaction properties
- Detection of regions of similarities and differences in molecular interaction fields (MIFs)
- Correlation of MIFs with functional parameters
- Interaction properties: electrostatic, hydrophobic etc. potentials
- Superimposed protein structures needed



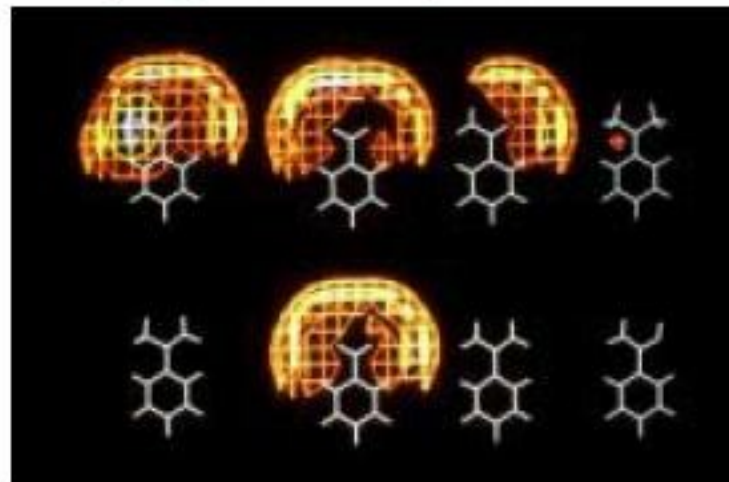
- Interaction fields are calculated on a set of points
- Field values on corresponding points are compared
- Φ = electrostatic potential, shape, probe interaction field, ...

similarity of small molecules

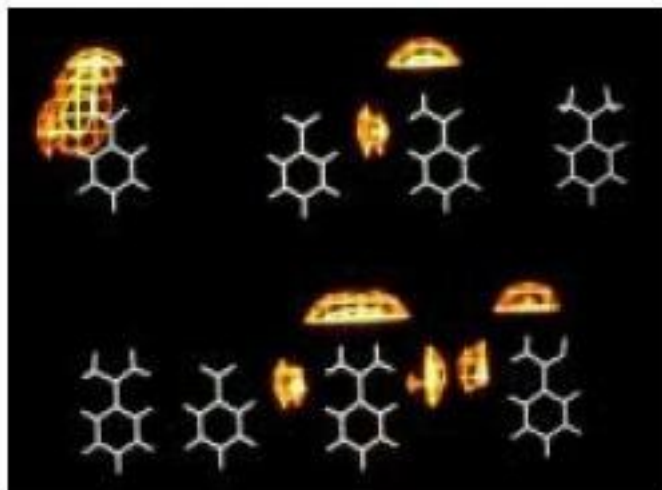
Similarity and Diversity



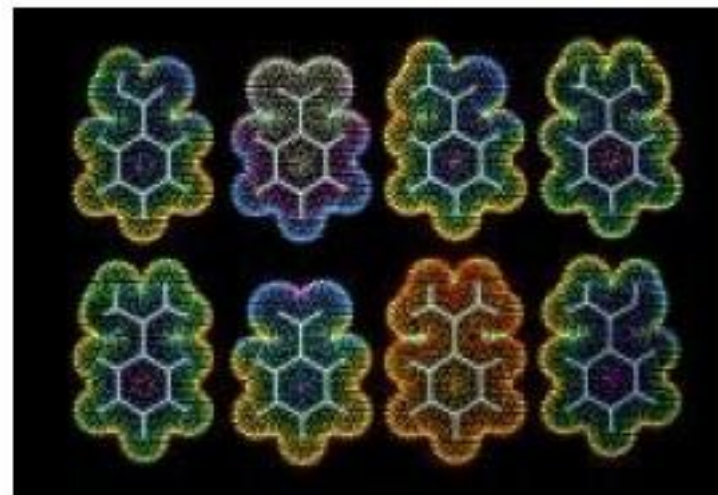
Hydrogen Bond Acceptor Potentials



Hydrogen Bond Donor Potentials



Volumes and Surface Potentials



Comparative Molecular Field Analysis (CoMFA)
Comparative Molecular Similarity Indices Analysis (ComSIA)

Similarity index (SI)

property X of A and B with N bits

- Euclidean

$$Euclidean_{AB} = \sqrt{\sum_{i=1}^N (X_{iA} - X_{iB})^2}$$

- Tanimoto

$$Tanimoto_{AB} = \frac{\sum_{i=1}^N X_{iA} X_{iB}}{\sum_{i=1}^N (X_{iA})^2 + \sum_{i=1}^N (X_{iB})^2 - \sum_{i=1}^N X_{iA} X_{iB}}$$

- Hodgkin

$$Hodgkin_{AB} = \frac{2\sum_{i=1}^N X_{iA} X_{iB}}{\sum_{i=1}^N (X_{iA})^2 + \sum_{i=1}^N (X_{iB})^2}$$

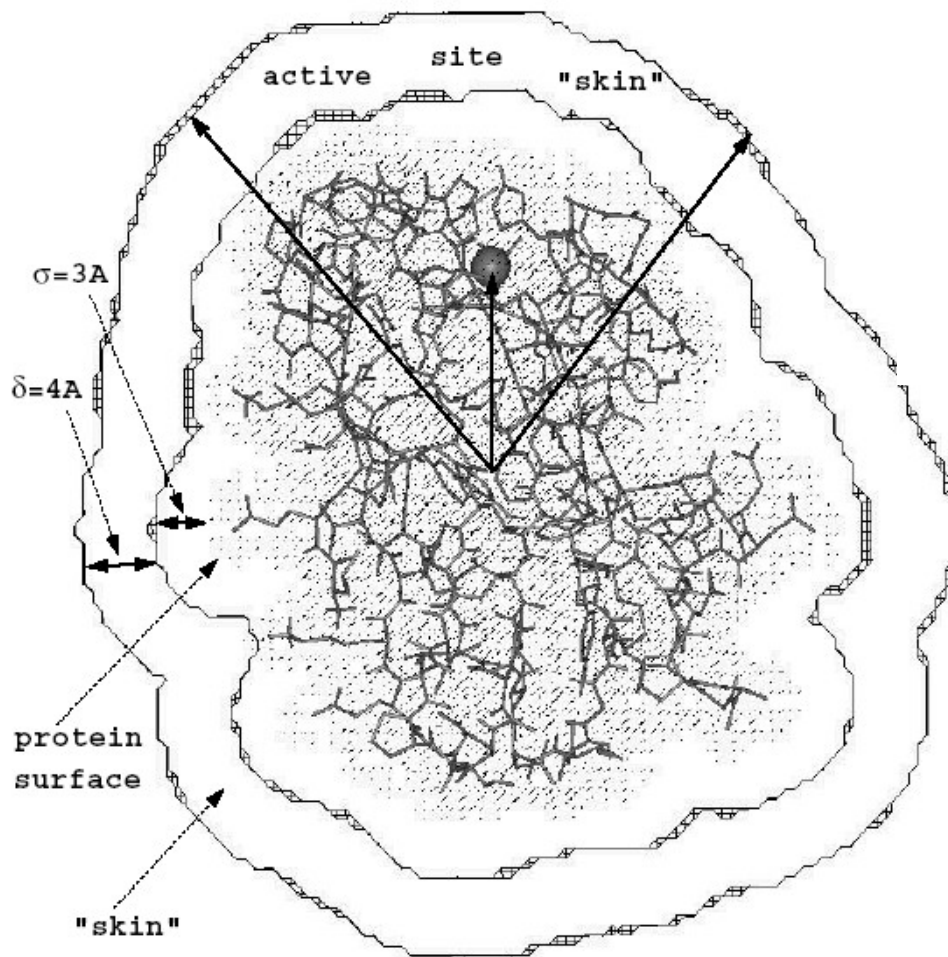
- Carbo

$$Carbo_{AB} = \frac{\sum_{i=1}^N X_{iA} X_{iB}}{\sqrt{\sum_{i=1}^N (X_{iA})^2 + \sum_{i=1}^N (X_{iB})^2}}$$

- electrostatic distance

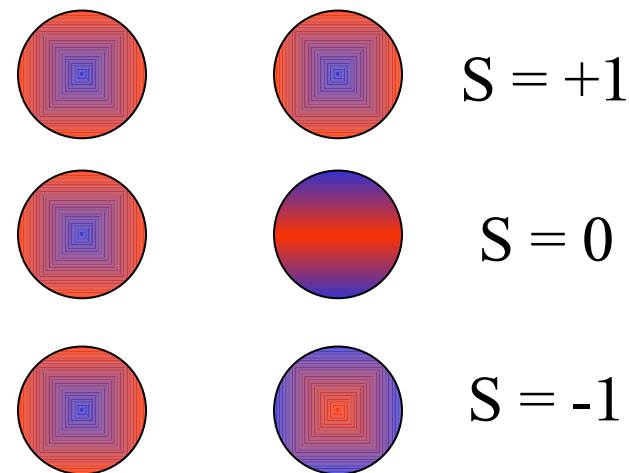
$$D_{ab} = \sqrt{1 - SI_{ab}}$$

PIPSA: Protein Interaction Property Similarity Analysis



$$SI_{12} = \frac{2(\mathbf{p}_1, \mathbf{p}_2)}{(\mathbf{p}_1, \mathbf{p}_1) + (\mathbf{p}_2, \mathbf{p}_2)}$$

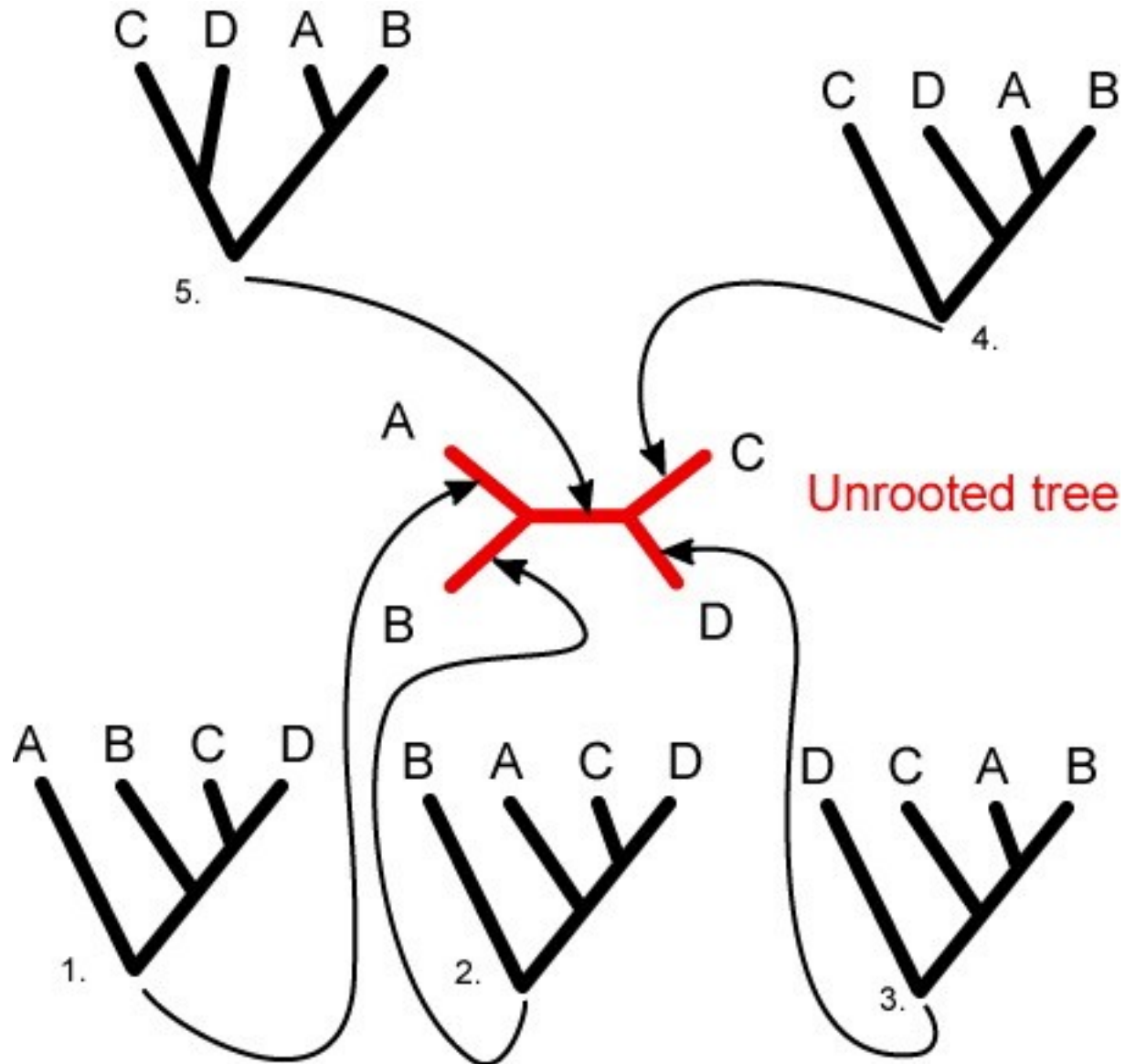
$$(\mathbf{p}_1, \mathbf{p}_2) = \sum_{i,j,k} \phi_1(i, j, k) \phi_2(i, j, k)$$



Wade et al., PNAS, 1998; Blomberg et al. Proteins 1999;

De Rienzo et al. Protein Sci. 2000; Wade et al. Intl. J. Quant. Chem. 2001

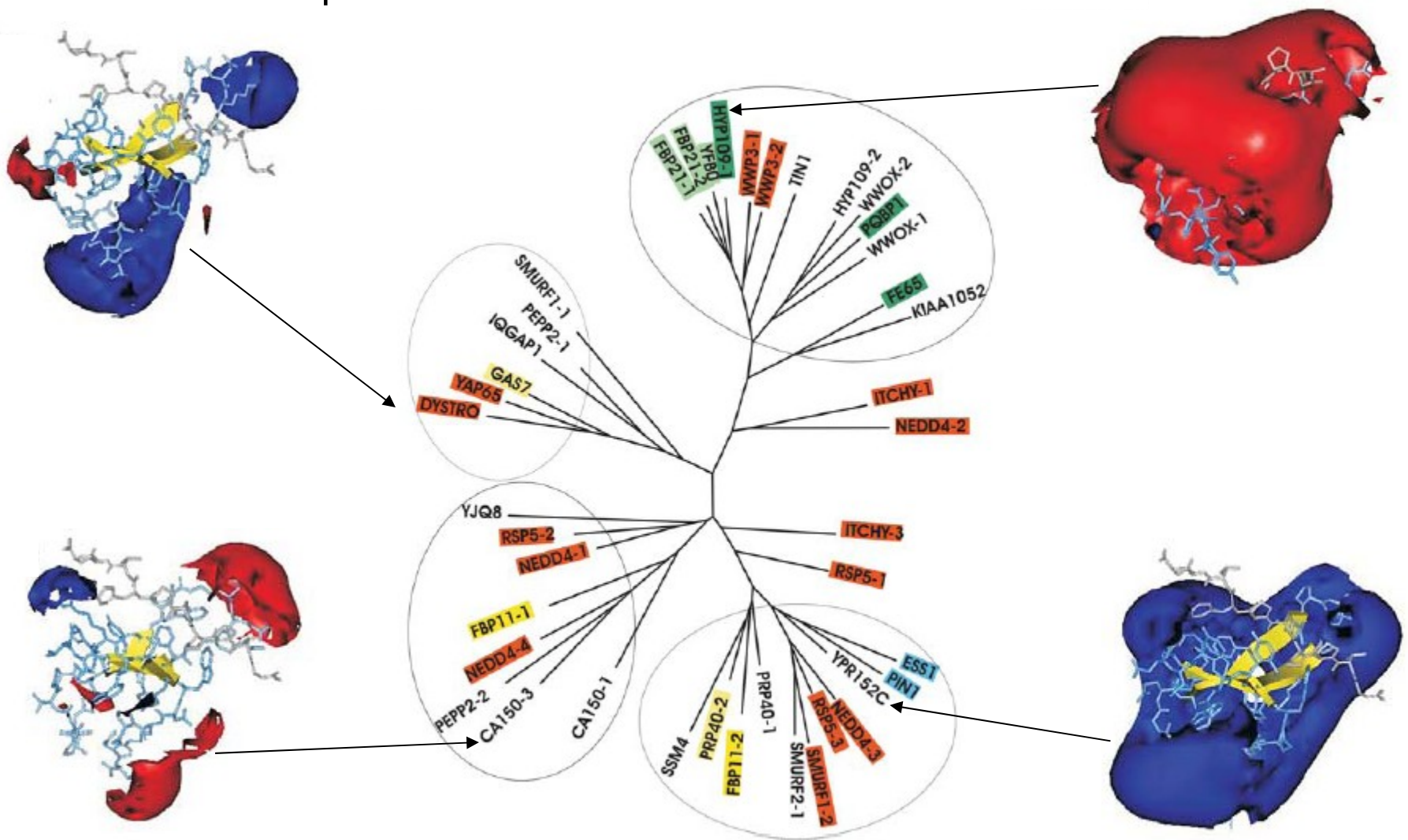
rooted and unrooted trees



42 WW Domains:

PIPSA epogram for Molecular electrostatic potential

isopotential contours: -0.4 / +0.4 kcal/mol/e

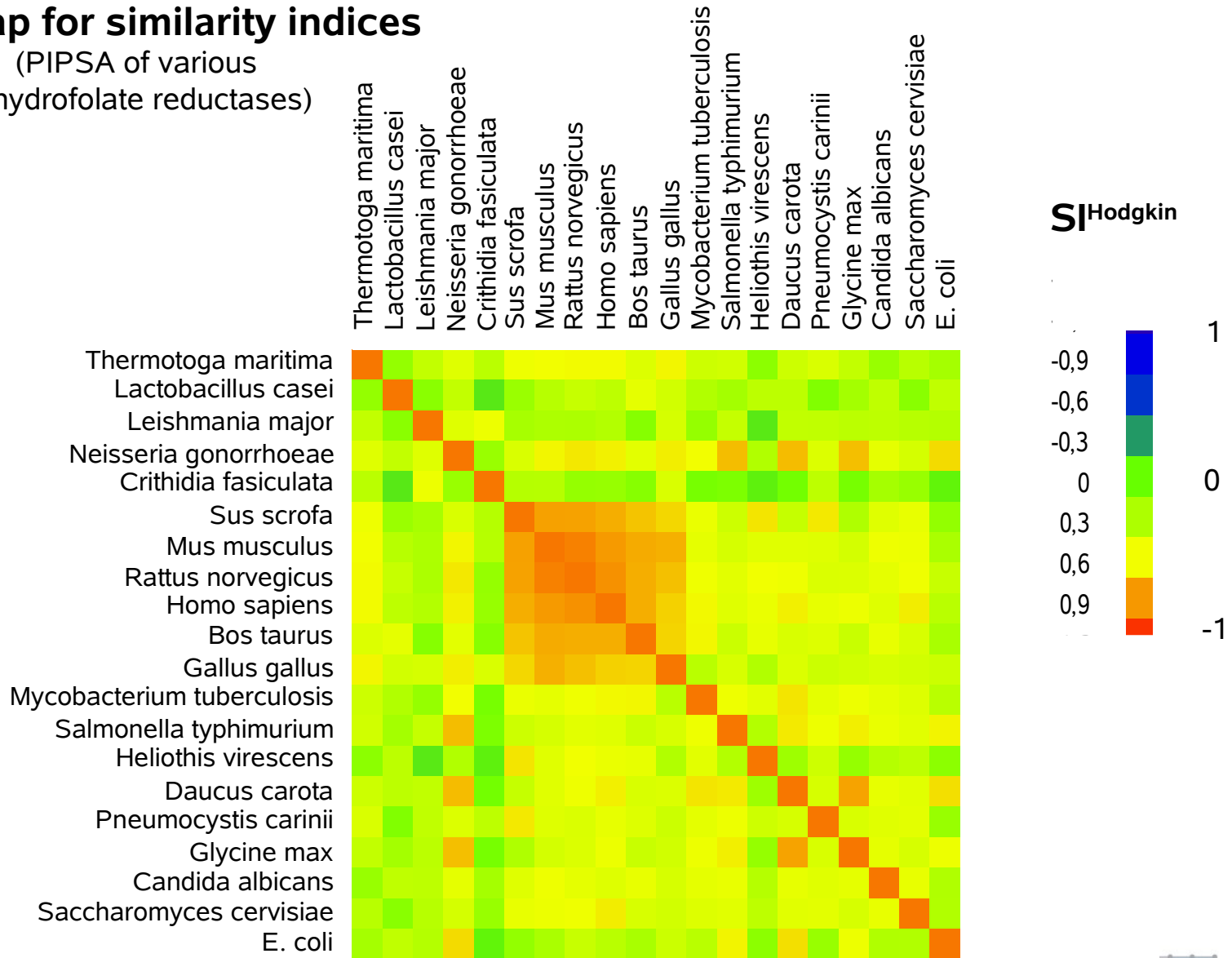


Schleinkofer, Wiedemann et al, JMB (2004) 344, 865-881

heat map

heat map for similarity indices

(PIPSA of various dihydrofolate reductases)

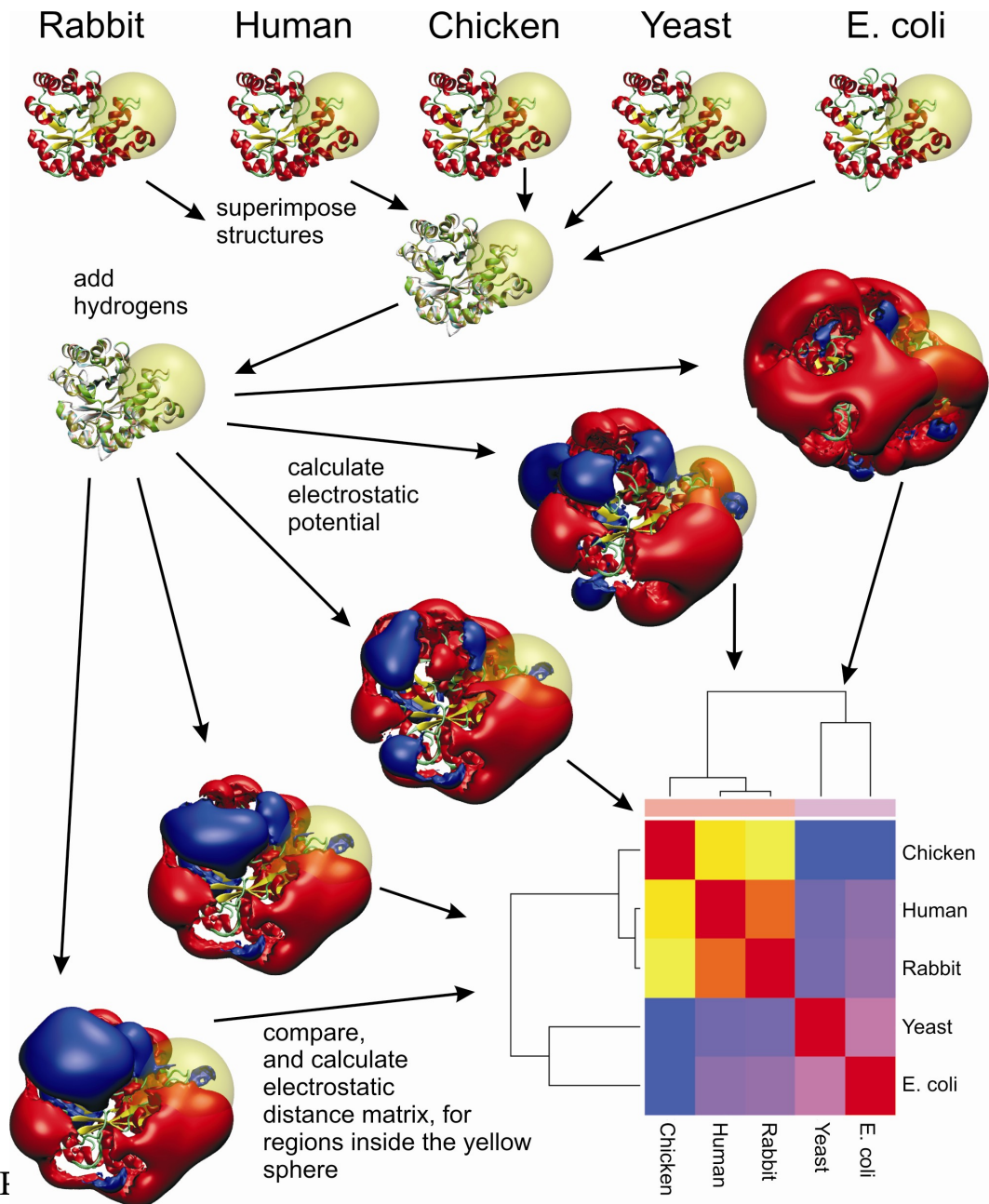


webPIPSA

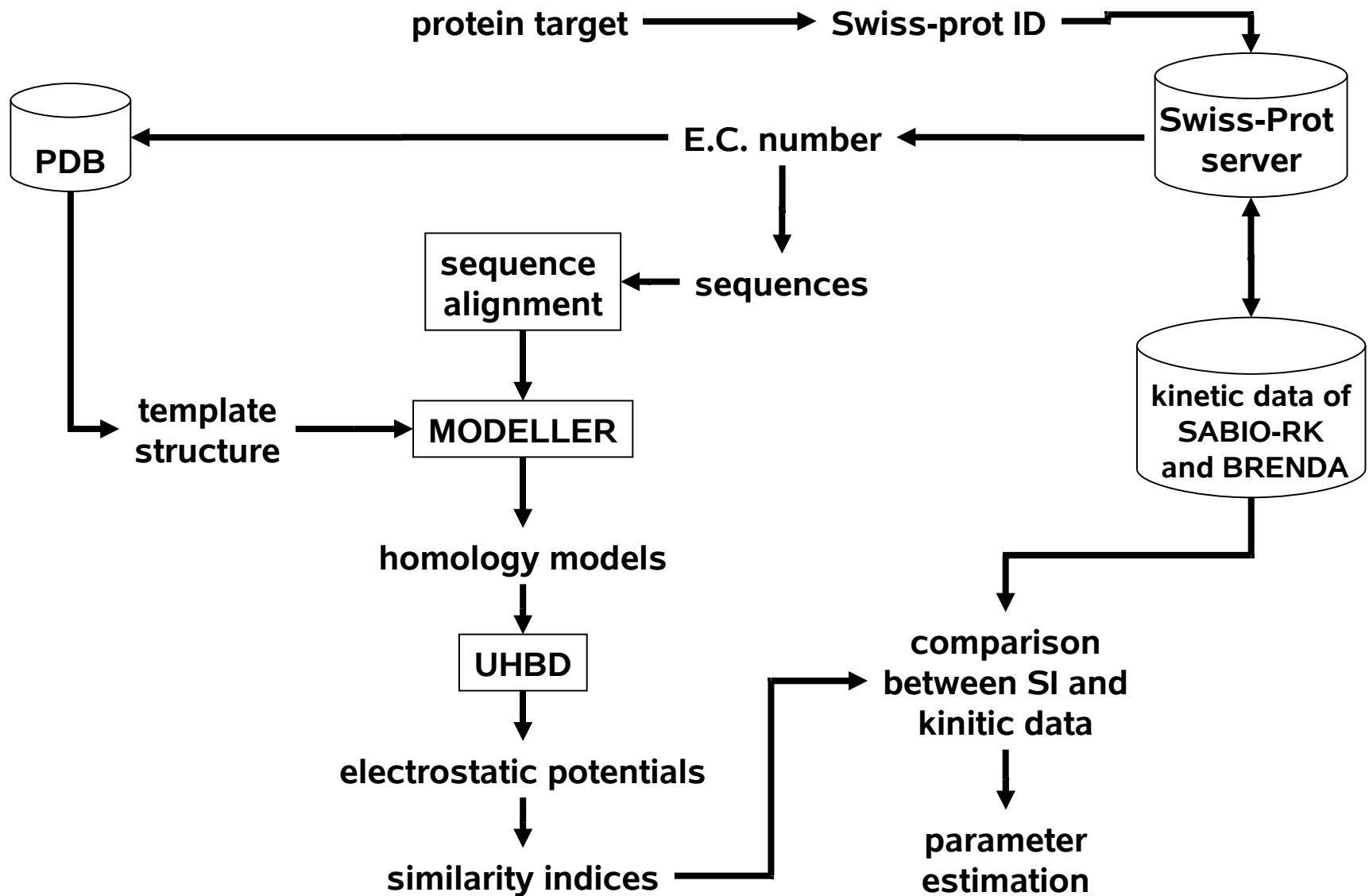
webPIPSA:
pipasa.embl.org

triosephosphate
 isomerase

Richter et al.,
Nucleic Acids Research, 2008



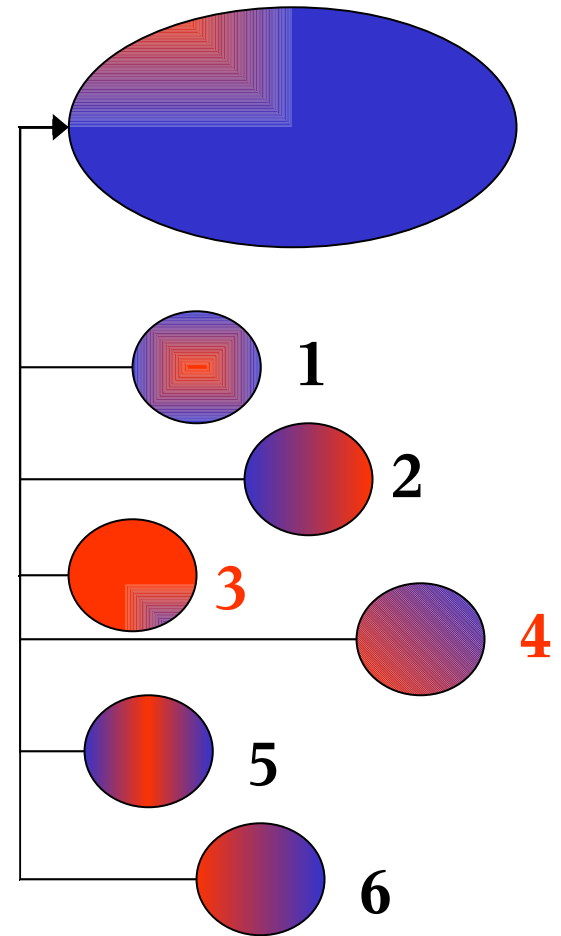
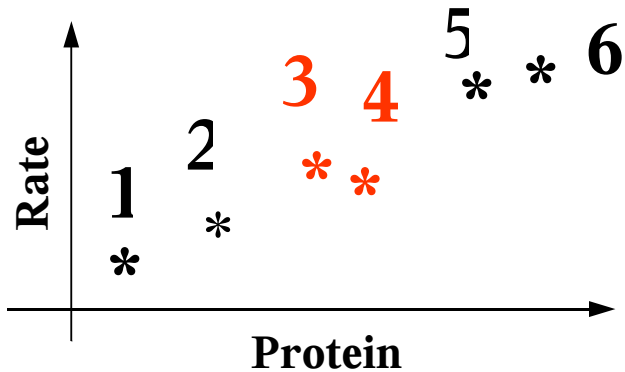
webPIPSA: workflow with homology modeling



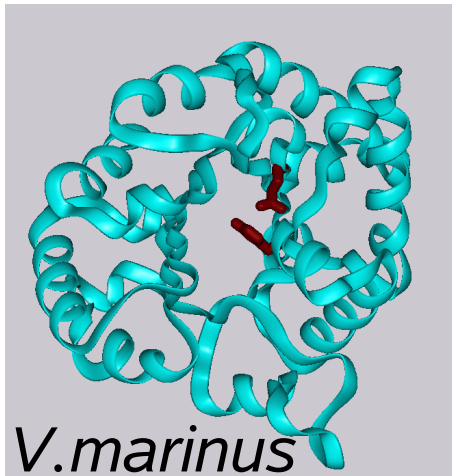
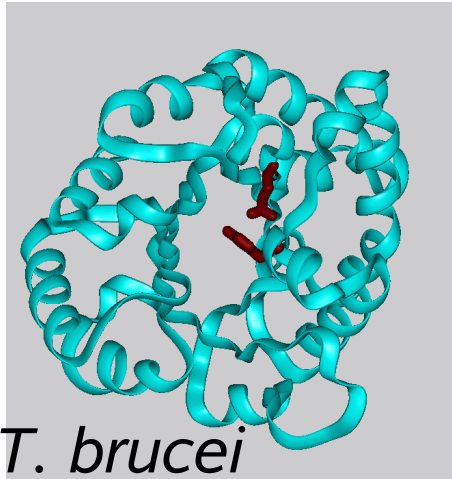
qPIPSA

Quantitative PIPSA (qPIPSA)

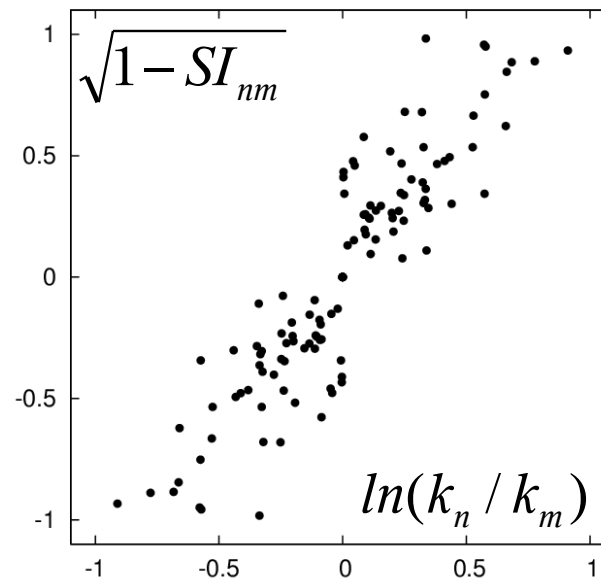
- Rates relative to one Partner
- Training Set Required with Experimental Information
- Predict Relative Ordering and Trends



triose phosphate isomerases from different species



- 40/55% sequence identity/homology
- same fold
- very similar active site
- factor of 3 difference in K_{cat}/K_m

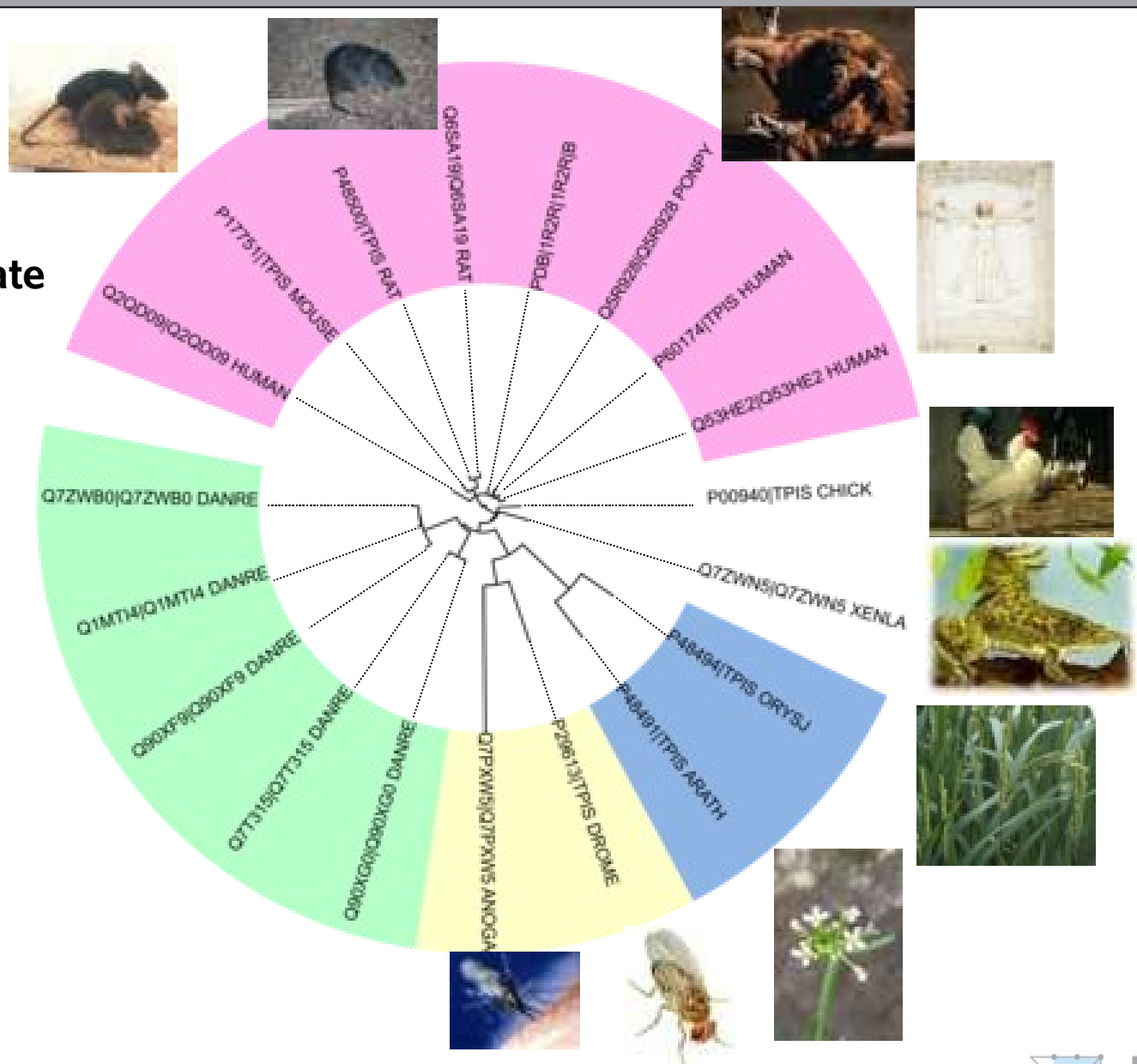


12 species:
 Giardia lamblia
 Spinach
 Chicken
 E. coli
 Human
 L. mexicana
 P. falciparum
 Rabbit
 T. brucei
 T. cruzi
 V. marinus
 Yeast

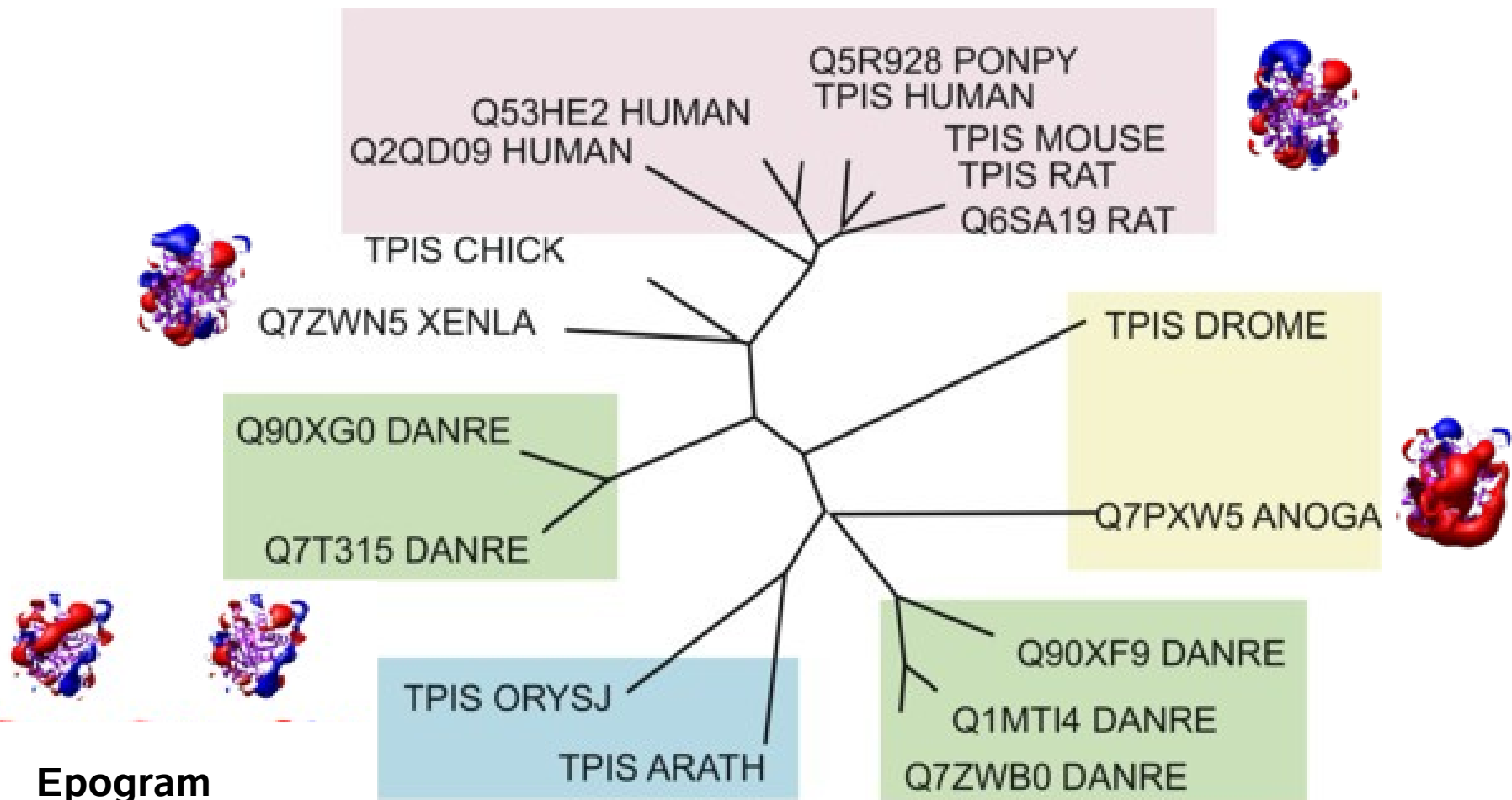
$$\ln(k_1/k_2) \approx \Delta G_2^\ddagger - \Delta G_1^\ddagger \approx \sqrt{\langle (\mathbf{p}_1 - \mathbf{p}_2)^2 \rangle} \approx \alpha \sqrt{(1 - SI_{12})}$$

Functional Annotation of Enzymes by PIPSA

Triosephosphate isomerases



Functional Annotation of Enzymes by PIPSA



Epogram

Tree-like diagram
based on
electrostatic potential

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€€€: Klaus Tschira Foundation, BMBF, DAAD, NIH, EU, DFG

MCM

<http://www.eml-research.de/english/research/mcm>
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PIPSA

webPIPSA:

<http://pipsa.eml.org>

<http://sycamore.eml.org>

download software:

<http://projects.villa-bosch.de/mcm/software/pipsa>

PIPSA

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