



#### Dissecting the bases of Bacillus thuringiensis host specificity: genomic and proteomic approaches

Yurv Malovichko<sup>12\*</sup>, Daria Gorbach<sup>2</sup>, Maria Belousova<sup>1</sup>, Ekaterina Romanovskava<sup>2</sup>, Elena Lukasheva<sup>2</sup>, Arseniy Lobov<sup>23</sup>, Christian Ihling<sup>4</sup>, Andrei Frolov<sup>24</sup>, Anton Nizhnikov<sup>12</sup>, Kirill Antonets<sup>12</sup>

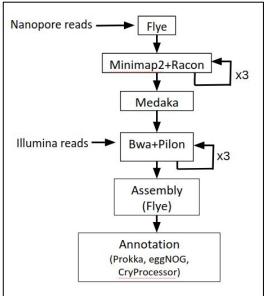
<sup>1</sup>Laboratory for Proteomics of SupraOrganismal Systems, All-Russian Research Institute of Agricultural Microbiology, Saint Petersburg, Russia <sup>2</sup>Department of Biology, St. Petersburg State University, Saint Petersburg, Russia <sup>3</sup>Laboratory of Regenerative Biomedicine, Institute of Cytology, Russian Academy of Sciences, Saint Petersburg, Russia <sup>4</sup>Department of Pharmaceutical Chemistry and Bioanalytics, Institute of Pharmacy, Martin-Luther Universität Halle-Wittenberg, Halle, Germany

**Background.** Bacillus thuringiensis (Bt) is a spore-forming bacterium affecting a wide range of invertebrate hosts. Bt is renowned for its high specificity towards the hosts; however, the mechanisms underlying this specificity remain unclear. In the present work we combine whole genome sequencing (WGS), qualitative and quantitative proteomics approaches to study the differences between four strains of three different serovars at vegetative and sporulating states and assess their virulence factors in respect to their host range.

**Table 1.** The list of strains used in this study. Vegetative cultures were grown on LB medium, and sporulating cultures grown on T3 medium for all four strains at 30°C for 72h

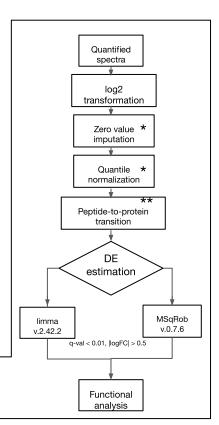
Strain	Serovar	Host range
109/25	darmstadiensis	Lepidoptera, Coleoptera, mites
800/15	thuringiensis	Coleoptera, Hemiptera
800/3	israeliensis	Diptera
800/3-15	israeliensis	Avirulent

**Acknowledgements.** Proteomic assays were supported by the Russian Foundation for Basic Research (grant No 20-316-70020) Genomic part of this work was previously accomplished under support of the Russian Science Foundation (grant No 18-76-00028).



**Figure 1.** Genome assembly and annotation pipeline

Figure 2. Quantitative proteomic analysis pipeline. Mass-spectrometry (MS) step was performed using HPLC/ESI-Orbitrap MS approach. \* Performed using MSnbase v.2.12.0 R package. \*\* Performed using MSqRob v.0.7.6 R package.



<sup>\*</sup> e-mail: yu.malovichko@arriam.ru

## Comparative genomics approach

Table 2. Genomes ON-based assembly statistics

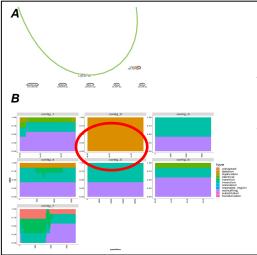
Strain	ON read number	Genome length, bp	N50, bp	Coverage, x
109/25	58289	273589223	9080	49.74349509
800/15	96747	443435819	8193	80.62469436
800/3	71602	365644310	9198	66.48078364
800/3-15	444850	863249530	4674	156.95446

**Table 3.** Genomes annotation summary

Strain	No of contigs	No of genes
109/25	10	6525
800/15	8	5951
800/3	7	6412
800/3-15	10	6307

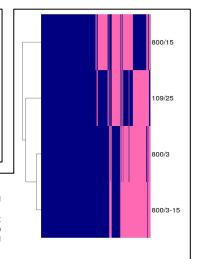
Table 4. Cry genes found with CryProcessor

Strain	Cry genes found
109/25	Cry1Ea10
800/15	Cry1Ba1 Cry1Ab12
800/3	Cry4Ba1 Cry4Aa2 Cry10Aa3
800/3-15	N/A



**Figure 3.** Results of genome assembly. **A.** A representation of strain 800/3 genome assembly. Linear contig represents chromosome, small circular contigs represent plasmids. **B.** A MUMmer plot of strain 800/3-15 genome aligned to genome of virulent ser. *israeliensis* strains. Highlighted contig absent in strain 800/3-15 bears 120 genes, of which 3 were found to encode for Cry toxins

Figure 4. A pan-genome reconstruction of the studied strains according to Roary. Blue lines indicate present orthologues, pink lines indicate absent ones. Similarity threshold was set to 90%



# CryProcessor (Shikov et al., 2020).

To mine for novel *Bt* insecticidal toxin genes, we elaborated a fast HMM-based tool, which can be found at the following link:

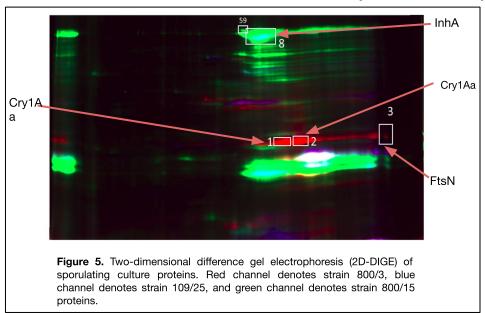


# Minor virulence factors of *Bt* (Malovichko *et al.*, 2019)

Currently we are working on a project dedicated to distribution of virulence factors genes unrelated to Cry toxins among *Bt* strains. A literature review on this topic and some preliminary results can be found at the following link:



## Qualitative/quantitative proteomics approach



**Table 5.** A total number of proteins identified from 59 differential spots using FPLC/ESI-quadrupole MS

Serovar	109/25	800/15	800/3
No of identified proteins	15	14	11

Quantitative proteomics assays were performed using a non-labeled protocol for HPLC/ESI-Orbitrap mass-spectrometry. Annotated and quantified spectra underwent MLE/MinDet imputation and quantile-dependent normalization with MSnBase. Differential production was assessed with MSqRob and limma packages at protein level with q-value threshold equal 0.01.

**Table 6.** Differential protein production in sporulating cultures (predictor) compared to vegetative cultures (intercept) according to MSqRob

Differentially produced protein, No		Strain			
	109/25	800/15	800/3	800/3-15	
Promoted	16	24	24	55	
Repressed	25	50	55	80	

**Table 7.** Differential protein production in sporulating cultures compared measured between the studied strains. In each pair the first strain indicates intercept (baseline) and the latter one indicate predictor

Differentially produced protein, No	Strain			
	109/25 vs 800/15	109/25 vs 800/3	800/15 vs 800/3	800/3 vs 800/3-15
Promoted	24	0	21	3
Repressed	16	0	24	8

Highlights. Several functional groups of proteins were enriched in the obtained datasets for interstrain comparisons. While for vegetative cells differentially produced proteins comprised mostly cellular metabolism enzymes, those differentially produced between sporulating cultures enclosed certain major virulence determinants (e.g. Cry and Cyt toxins and InhA/ColB metalloproteases) as well as exosporium maturation proteins. These assumptions, however, need further elucidation with the statistically correct assessment of Gene Ontology and/or KEGG term representation. Taken together, the presented data clearly reflect the differences between the strains studied, but their association with the observed phenotypes and host range is yet to be accomplished.